

## Enhanced root carbon allocation through organic farming is restricted to topsoils

**Running title:** Drivers of root biomass in farming systems

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1 **Abstract**

2           Soils store significant amounts of carbon (C) and thus can play a critical role for mitigating  
3 climate change. Crop roots represent the main C source in agricultural soils and are particularly  
4 important for long-term C storage in agroecosystems. To evaluate the potential of different farming  
5 systems to contribute to soil C sequestration and thus climate change mitigation, it is of great importance  
6 to gain a better understanding of the factors influencing root C allocation and distribution. So far, it is  
7 still unclear how root C allocation varies among farming systems and whether the choice of management  
8 practices can help to enhance root C inputs. In this study, we compared root C allocation in three main  
9 arable farming systems, namely organic, no-till, and conventional farming. We assessed root biomass,  
10 vertical root distribution to 0.75 m soil depth, and root-shoot ratios in 24 winter wheat fields. We further  
11 evaluated the relative importance of the farming system compared to site conditions and quantified the  
12 contribution of individual management practices and pedoclimatic drivers. Farming system explained  
13 one third of the variation in topsoil root biomass and root-shoot ratios, both being strongly positively  
14 related to weed biomass and soil organic C content and negatively to mineral nitrogen fertilization  
15 intensity. Root C allocation was significantly higher in organic farming as illustrated by an increase in  
16 root biomass (+40%) and root-shoot ratios (+60%) compared to conventional farming. By contrast, the  
17 overall impact of no-till was low. The importance of pedoclimatic conditions increased substantially  
18 with soil depth and deep root biomass was largely controlled by precipitation and soil texture, while the  
19 impact of management was close to zero. Our findings highlight the potential of organic farming in  
20 promoting root C inputs to topsoils and thereby contributing to soil organic matter build-up and  
21 improved soil quality in agroecosystems.

22 **Keywords**

23 root carbon inputs; farming system; agricultural management; on-farm study; root biomass distribution;  
24 subsoil

## 25 1. Introduction

26 Soils play a prominent role in the global carbon (C) cycle as they contain substantially more C  
27 than the atmosphere and land vegetation combined (Lehmann and Kleber, 2015). Increasing soil organic  
28 C therefore holds great promise for mitigating climate change. Agricultural soils could be a key in this  
29 effort because 34% of the land surface is currently under agricultural use (Ritchie and Roser, 2020) and  
30 management substantially influences soil organic C storage by altering inputs and decomposition rates  
31 (Janzen, 2015; Paustian *et al.*, 2016).

32 Root C is one of the most important contributors to soil organic C and constitutes up to 90% of  
33 all C inputs to arable soils (Kätterer *et al.*, 2011). Due to its resistant chemical composition (Rasse *et*  
34 *al.*, 2005) and preferential incorporation into more stable fractions (Ghafoor *et al.*, 2017), root C has a  
35 longer residence time in soil than C derived from above ground crop residues and manure (Kätterer *et*  
36 *al.*, 2011; Menichetti *et al.*, 2015; Zhang *et al.*, 2015). Particularly, root C inputs to deep soil have been  
37 linked to long-term C storage (Russell *et al.*, 2009; Fan *et al.*, 2019) due to the low decomposer  
38 abundance and high storage capacity of deep unsaturated layers (Rasse *et al.*, 2005; Rumpel *et al.*, 2012;  
39 Sanaullah *et al.*, 2016). Hence, the promotion of more and deeper roots has been proposed as a strategy  
40 to mitigate climate change with an estimated potential to remove atmospheric CO<sub>2</sub> of about 1 Pg yr<sup>-1</sup>  
41 (Lynch and Wojciechowski, 2015; Paustian *et al.*, 2016; Pierret *et al.*, 2016). Thus, it is crucial to  
42 understand how management can promote root C inputs to agricultural soils in order to sequester C in  
43 the long-term, but also to stimulate C dynamics, thereby enhancing the manifold benefits of soil organic  
44 matter for agricultural soils (Janzen, 2015; Paustian *et al.*, 2016).

45 Agricultural management affects root biomass allocation in various ways by its impact on crop  
46 nutrition and soil properties through e.g. type and amount of fertilization, crop rotation, or soil tillage  
47 (Malhi and Lemke, 2007; Chirinda *et al.*, 2012; Qin *et al.*, 2018). For instance, in organic farming, the  
48 application of synthetic nutrient inputs is prohibited, which often leads to reduced mineral nitrogen (N)  
49 availability (Lorenz and Lal, 2016). It is expected that this increases biomass allocation below ground  
50 as crops need to cope with primarily growth-limiting resources (Lynch *et al.*, 2012; Poorter *et al.*, 2012).  
51 No-till farming is another alternative to conventional farming and is characterised by reduced or zero  
52 soil disturbance through tillage. Hence, it often results in accumulation of organic matter and nutrients

53 but also increased bulk density in the topsoil (Huggins and Reganold, 2008; Powlson *et al.*, 2014). This  
54 may lead to a shift in biomass allocation and increased superficial root proliferation (Qin *et al.*, 2018;  
55 Mondal *et al.*, 2020), thereby altering vertical root distribution (Dwyer *et al.*, 1996; Ball-Coelho *et al.*,  
56 1998; Barzegar *et al.*, 2004). So far, the influence of different farming systems on root C allocation has  
57 still not been clearly established and current knowledge is based on controlled field studies conducted  
58 at a small number of sites. In organic farming, both similar (Steingrobe *et al.*, 2001; Lazicki *et al.*, 2016;  
59 Hirte *et al.*, 2018a) and higher (Chirinda *et al.*, 2012; Hu *et al.*, 2018) root biomass compared to  
60 conventional farming has been reported for cereals. No-till was even found to influence root biomass in  
61 any direction for cereals or rapeseed, i.e. tillage effects were negative, absent, or positive (Plaza-Bonilla  
62 *et al.*, 2014; Li *et al.*, 2017; Sarker *et al.*, 2017).

63 The unclear picture of how agricultural management influences root C allocation may be linked  
64 to the impact of soil and climate characteristics that often overlay management effects. Soil properties  
65 such as mechanical impedance or nutrient availability as well as climatic conditions such as precipitation  
66 or temperature affect root growth to a large extent and complex interactions of stimuli often obliterate  
67 root response to individual drivers (reviewed by Rich and Watt, 2013). Consequently, biomass  
68 allocation to roots and shoots can vary by a factor of 10 across environments (Enquist and Niklas, 2002;  
69 Poorter *et al.*, 2012). In order to unravel the potential of agricultural management to enhance root C  
70 inputs to soil (Paustian *et al.*, 2016; Dignac *et al.*, 2017), management effects need to be assessed over  
71 a wide range of pedoclimatic conditions. On-farm measurements over multiple locations can not only  
72 provide practice-related, generalizable results but could also allow for quantitative comparisons of the  
73 effects of specific management practices on crop parameters beyond classified farming systems  
74 (Nkurunziza *et al.*, 2017; Büchi *et al.*, 2019).

75 We therefore established a network of 24 farms classified as conventional, no-till, or organic in  
76 Switzerland and investigated root biomass in the top and subsoil in winter wheat fields. In addition, we  
77 collected detailed information on management practices and soil and climate conditions for each field.  
78 Our objectives were (i) to assess the impact of organic, conventional and no-till farming on root biomass  
79 and plant biomass allocation and (ii) to evaluate the relative importance of management- and site-related  
80 variables for root and shoot biomass, root-shoot ratios, and vertical root distribution.

## 81 2. Methods

### 82 2.1. Farming systems and sites

83 The study was conducted in 2016 on 24 commercial farms in the northern part of Switzerland,  
84 which were categorized as conventional with tillage (conventional), conventional without tillage (no-  
85 till), or organic with tillage (organic) according to the farm structure census 2015 (Supplementary table  
86 1; FSO, 2017; Büchi *et al.*, 2019). No-till soil management implied that not more than 25% of the soil  
87 surface could be disturbed at sowing (Swiss Federal Council, 2013). All farms were managed according  
88 to the certification scheme Proof of Ecological Performance PEP (Swiss Federal Council, 2013), the  
89 guidelines of the Swiss Farmer Association for Integrated Production IP-Suisse (IP-SUISSE, 2019), or  
90 the regulations of the Federation of Swiss Organic Farmers BIO-Suisse (Swiss Federal Council, 1997).  
91 The farms were located at eight sites spread over a distance of roughly 100 km arranged in farming  
92 system triplets of one conventional, no-till, and organic farm each (Supplementary figure 1). The nearest  
93 weather stations operated by the Federal Office of Meteorology and Climatology with recorded long-  
94 term precipitation data were chosen as reference points for the sites (Supplementary table 2). Annual  
95 temperature and precipitation (1981–2010) for Zurich-Affoltern (08°31'04", 47°25'40"), which is  
96 centrally located within the study area, are 9.4 °C and 1054 mm, respectively.

### 97 2.2. Growth conditions of winter wheat

98 On each farm, one field was selected for plant and soil analyses. Winter wheat (*Triticum*  
99 *aestivum*, L.) was sown between 2 and 26 October 2015 and harvested between 18 July and 4 August  
100 2016. Varieties, type of fertilization, weed and pathogen control, and use of growth retardants differed  
101 between farms (Supplementary table 1). Organic fertilizers were applied as cattle or pig slurry using an  
102 injector or as cattle manure, compost, humus acid suspension, or granulated organic N fertilizer (Büchi  
103 *et al.*, 2019).

### 104 2.3. Root and shoot sampling

105 Root and shoot biomass of wheat and weeds was sampled at wheat flowering between 14 and  
106 23 June 2016. A circular area with a radius of 10 m and a distance of at least 20 m to the nearest edge  
107 of the field was defined as sampling area and divided into four quarters (Supplementary figure 2). Within

108 each quarter, shoot samples were taken directly above the ground on one randomly selected sampling  
109 plot covering four wheat rows of 0.5 m length with electric grass clippers and separated into wheat and  
110 weed shoot biomass. On the same sampling plots, root samples were collected by taking two soil cores,  
111 one within and one half-way between wheat rows, to a depth of 0.75 m by means of a metal sampling  
112 rod (inner diameter: 60 mm; lined with polyethylene film) driven into soil with an electric breaker  
113 (EH50, Wacker, Germany) and extracted with a 3-cylinder-lifting unit (ZGM-9E ECO, Nordmeyer  
114 Geotool GmbH, Germany). The cores were separated into three layers of 0.25 m length (top: 0–0.25 m,  
115 intermediate: 0.25–0.5 m, deep: 0.5–0.75 m) and stored in polyethylene film at 4 °C for a maximum of  
116 three weeks until further processing.

#### 117 2.4. Biomass determination

118 Roots were extracted from each soil core separately using an automated root washer  
119 (Hydropneumatic Elutriation System GVF 13000, Gillison`s Variety Fabrication Inc., USA). The field-  
120 fresh soil was dispersed for 10 minutes in a high-energy hydrovortex at a water pressure of  
121 approximately 350 kPa and roots were separated from the mineral fraction by flotation and recovered  
122 on a 0.5 mm mesh (Smucker et al., 1982). The thus retained root samples were transferred to aluminium  
123 dishes and extraneous organic matter was visually identified based on shape, structure, colour, and  
124 elasticity of particles and removed from the samples using tweezers (Schuurman and Goedewaagen,  
125 1971; Hirte et al., 2017). Identifiable weed roots, e.g. tap or rhizomatous roots, were removed from the  
126 root samples. However, a certain proportion of weed roots could not be distinguished from wheat roots  
127 by eye and remained in the samples. All plant material was dried at 55 °C until constant weight (shoots:  
128 72 h; roots: 48 h) and dry weight was recorded.

#### 129 2.5. Management and pedoclimatic variables

130 The following variables and their importance for root biomass and distribution were  
131 investigated: mineral N fertilization intensity, sowing density, above ground weed biomass, soil bulk  
132 density, soil texture, soil organic C, total N and available P in soil, and precipitation (Supplementary  
133 table 3). Mineral N fertilization intensity and sowing density were derived from questionnaires returned  
134 by the farmers (Büchi et al., 2019). Mineral N fertilization intensity was calculated from fertilizer-N

135 input (total N in mineral fertilizers and ammonium-N in organic fertilizers as estimated by Büchi *et al.*,  
136 2019) in the wheat season 2015/2016 as the amount of applied N ( $\text{kg ha}^{-1} \text{ season}^{-1}$ ) relative to the  
137 recommended amount of available N ( $\text{kg ha}^{-1} \text{ season}^{-1}$ ) for wheat according to the Principles of  
138 Agricultural Crop Fertilisation in Switzerland (Richner and Sinaj, 2017). Although wheat variety was  
139 an important aspect of management, this categorical information could not be accounted for due to the  
140 great diversity of 15 different genotypes and, thus, the lack of replications across fields (Supplementary  
141 table 1).

142 Soil was sampled on each farm between 20 April and 27 May 2016 for determination of soil  
143 texture, organic C, total N, available P, and bulk density. Except for bulk density, 15–20 samples were  
144 taken in five soil layers (0–0.05 m, 0.05–0.2 m, 0.2–0.25 m, 0.25–0.5 m, 0.5–0.75 m) on transect lines  
145 that ran in 45° angles to the seedling rows and divided the quarters for root and shoot biomass sampling.  
146 Composite samples per layer were dried and soil texture (sedimentation), organic C (oxidation with  
147 potassium dichromate), and available P ( $\text{CO}_2$ -saturated water extraction and colorimetry) were  
148 determined on 2-mm sieved fine soil according to the Swiss reference methods (Agroscope, 1996). Total  
149 soil N was measured after dry oxidation according to the Dumas method (Bremner, 1965). For soil bulk  
150 density measurements, undisturbed samples of 100 ml volume and 50 mm height were taken in the  
151 middle of each layer except the 0.5–0.75 m layer and oven-dried at 105°C for at least 72 h (Colombi *et*  
152 *al.*, 2019). Bulk density values of the 0.25–0.5 m layer were used for the 0.5–0.75 m layer. The weighted  
153 averages of variables measured on samples from the upper three layers (0–0.05 m, 0.05–0.2 m, 0.2–0.25  
154 m) served as composite values for the 0–0.25 m layer for further analyses.

155 Precipitation during the wheat growing season (October 2015 to June 2016) was retrieved from  
156 the nearest local weather station to each farm operated by either MeteoSwiss, the Federal Roads Office,  
157 the Cantons of Lucerne, Thurgovia, or Zurich, or MeteoGroup Switzerland. Due to clustering of farms  
158 within sites and limited spatial distribution of local weather stations, 12 data sets for the total of 24 farms  
159 were available. We tested the effect of cumulative precipitation during several time periods on the  
160 investigated response variables and found the strongest effect for precipitation between March and mid-  
161 June, i.e. between tillering and flowering, corresponding to the main part of the vegetative growth phase.  
162 From here on, we refer to this time period when we report values and the effect of precipitation.

163 2.6. Calculations and statistics

164 To extrapolate to field scale, root biomass sampled within and between rows was weighted with  
165 respect to row width for each layer individually (adapted from Frasier et al., 2016):

$$RB_{within} = \frac{M_{within}}{\pi * (\frac{D}{2})^2} * \frac{D}{s} \quad (1)$$

$$RB_{between} = \frac{M_{between}}{\pi * (\frac{D}{2})^2} * \frac{(s - D)}{s} \quad (2)$$

166 where  $RB_{within}$  and  $RB_{between}$  are root biomass ( $\text{g m}^{-2}$ ) within and between rows, respectively,  
167  $M_{within}$  and  $M_{between}$  are the dry weights of roots (g) extracted from the soil cores taken within and between  
168 rows, respectively,  $D$  is the inner diameter of the sampling rod (m), and  $s$  is the distance between rows  
169 (m). Root biomass was obtained by summing  $RB_{within}$  and  $RB_{between}$ . Root-shoot ratios were calculated  
170 for each subplot from averaged total root (0–0.75 m) and shoot biomass and were ln-transformed prior  
171 to statistical analysis (Poorter and Sack, 2012). Unless otherwise stated, root-shoot ratios relate to wheat  
172 shoot biomass (excluding weed) but were also analysed for wheat plus weed shoot biomass.

173 A few data points (12 out of 576) needed to be eliminated when problems with sampling or  
174 sample processing occurred (e.g. sieve clogging and root loss in the root washer). Consequently, root  
175 biomass could not be estimated for those instances and only 3 out of 4 field replications were used. Root  
176 and shoot biomass and root-shoot ratios of individual subplots on each farm were treated as lower-level  
177 replicates for statistical analysis and were averaged per farm for data presentation. Mean data for farming  
178 systems and sites are presented as averages of farming system/site and farm and standard errors of  
179 farming system/site.

180 We analysed the data in a three-step procedure and thereby investigated the following response  
181 variables: root biomass and the proportion of root biomass in the individual layers (0–0.25 m, 0.25–0.5  
182 m, 0.5–0.75 m) and total root biomass (0–0.75 m) of wheat and weeds, wheat shoot biomass, and root-  
183 shoot ratio. (i) To test for differences in response variables between farming systems and sites, we fitted



184 the data to mixed effects models (fixed factors: farming system and site; random factor: farm) and  
185 determined differences between group means by ANOVA and subsequent simultaneous multiple  
186 comparison of estimated marginal means of group pairs with Tukey-adjustment of p-values. (ii) To  
187 further evaluate the effects of the management and pedoclimatic variables on the response variables, we  
188 used mixed effects models (fixed factor: management or pedoclimatic variable; random factor: farm) in  
189 univariate analyses and ANOVA. (iii) To determine the relative importance of (a) farming system and  
190 site and (b) management and pedoclimatic variables for the response variables, we conducted  
191 multivariate linear regressions without prior variable selection and calculated variance decomposition  
192 metrics: (a) LMG metrics for uncorrelated categorical regressors (Lindeman Merenda Gold; Lindeman,  
193 1980) and (b) CAR scores for correlated numerical regressors (Correlation-Adjusted coRrelation; Zuber  
194 and Strimmer, 2011). While LMG metrics are unweighted averages over orderings of sequential  
195 contributions of explanatory variables to models of different sizes (Grömping, 2015), CAR scores are  
196 based on simultaneous orthogonalization of correlated explanatory variables and subsequent estimation  
197 of marginal correlations between response and decorrelated explanatory variables (Zuber and Strimmer,  
198 2011). Shoot biomass and root-shoot ratios were related to soil variables in the top layer. We considered  
199 a significance level of  $p < 0.05$ .

200 We used the software R version 3.4.2 (R Core Team, 2019) and the R packages lme4 (Bates et  
201 al., 2015), lmerTest (Kuznetsova et al., 2017), pbkrtest (Halekoh and Højsgaard, 2014), emmeans  
202 (Lenth, 2018), and relaimpo (Grömping and Lehrkamp, 2018) for statistical analyses and the R packages  
203 ggplot2 (Wickham, 2016), GGally (Schloerke et al., 2018), gridExtra (Auguie, 2017), and lemon  
204 (Edwards, 2019) for data visualization.

### 205 3. Results

206 We analysed total root biomass, vertical root distribution, wheat shoot biomass, and root shoot  
207 ratios from 24 farms arranged in farming system triplets (conventional, no-till, organic) that were located  
208 at eight sites in Switzerland. The sites spread over a distance of just 100 km, yet pedoclimatic  
209 characteristics varied considerably among farms (Supplementary table 3). Total root biomass in the 0–  
210 0.75 m soil profile ranged among individual farms from 87–274 g m<sup>-2</sup>. Root biomass varied between

211 55–178 g m<sup>-2</sup> in the top layer, 12–53 g m<sup>-2</sup> in the intermediate layer, and 7–43 g m<sup>-2</sup> in the deep layer,  
212 corresponding to 55–78%, 10–28%, and 8–22% in the respective layers of total root biomass. Wheat  
213 shoot biomass ranged among farms from 909–1692 g m<sup>-2</sup> and root-shoot ratios from 0.07–0.22.

### 214 3.1. Differences in root parameters between farming systems

215 Total root biomass was 132 g m<sup>-2</sup> in conventional, 156 g m<sup>-2</sup> in no-till, and 182 g m<sup>-2</sup> in organic  
216 farming and was significantly higher in organic than conventional ( $p = 0.018$ ) and intermediate in no-  
217 till farming (Figure 1). Differences between farming systems were limited to the top layer, where root  
218 biomass was 87, 101, and 132 g m<sup>-2</sup> in conventional, no-till, and organic farming, respectively, and  
219 significantly higher in organic compared to both conventional ( $p = 0.003$ ) and no-till farming ( $p = 0.032$ ;  
220 Figure 1). The proportion of topsoil root biomass was highest in organic (73%), lowest in no-till (64%;  
221  $p = 0.017$ ), and intermediate in conventional farming (66%; Supplementary figure 2). In the intermediate  
222 and deep layer, respectively, root biomass and its proportion were similar among farming systems,  
223 averaging 27 g m<sup>-2</sup> (18%) and 23 g m<sup>-2</sup> (14%; Figure 1; Supplementary figure 2).

224 Wheat shoot biomass at flowering was similar among farming systems and averaged 1311 g m<sup>-2</sup>  
225 (Figure 1). Consequently, root-shoot ratios were significantly higher in organic farming than in both  
226 conventional and no-till farming, irrespective of whether shoot biomass referred to wheat shoot biomass  
227 only (organic 0.15; conventional 0.09,  $p < 0.001$ ; no-till 0.11,  $p = 0.002$ ) or wheat plus weed shoot  
228 biomass (organic 0.14; conventional 0.09,  $p < 0.001$ ; no-till 0.10,  $p = 0.005$ ; Figure 1).

### 229 3.2. Variation in root parameters among sites

230 Total root biomass ranged from 105–221 g m<sup>-2</sup> among the eight farming system triplets and  
231 differed significantly between sites ( $p = 0.011$ ). In addition to the large variation in topsoil root biomass  
232 (75–151 g m<sup>-2</sup>;  $p = 0.015$ ), significant differences between sites also occurred in deep root biomass (11–  
233 35 g m<sup>-2</sup>;  $p = 0.014$ ), while root biomass was similar in the intermediate layer (27 g m<sup>-2</sup>). Vertical root  
234 distribution was not significantly affected by site conditions as the proportion of root biomass was  
235 similar among sites in all layers (top: 68%, intermediate: 18%, deep: 15%). Similar to the farming  
236 system comparison, wheat shoot biomass at flowering was similar among sites (1311 g m<sup>-2</sup>) but root-  
237 shoot ratios differed significantly (0.07–0.18;  $p < 0.001$ ; Supplementary table 4).

238 3.3. Differences in management and pedoclimatic variables between farming systems and sites

239 Compared to conventional and no-till farming, organic farming involved lower N fertilization  
240 intensity ( $p = 0.003$  and  $0.025$ , respectively) but higher weed biomass ( $p = 0.011$  and  $0.009$ , respectively;  
241 Supplementary table 3). Topsoil bulk density was higher in no-till than in conventional and organic  
242 farming ( $p < 0.001$  each). All other variables were similar among farming systems except for organic C  
243 and total N in the intermediate layer, which were higher in organic than in no-till ( $p = 0.009$  and  $0.017$ ,  
244 respectively) and intermediate in conventional farming (Supplementary table 3). The sites differed in  
245 mineral N fertilization intensity, topsoil bulk density, precipitation, soil organic C, total soil N, sand,  
246 silt, and clay content in the top and intermediate layer (see Supplementary table 3 for p-values). In the  
247 deep layer, all soil variables were similar among both farming systems and sites (data not shown).

248 3.4. Explained variation in root and shoot biomass and root-shoot ratio

249 *Farming system and site*

250 Farming system and site as explanatory variables accounted for 19 and 54%, respectively, of  
251 the variation in total root biomass. In the top, intermediate, and deep layer, respectively, the variation in  
252 root biomass was by 32, 11, and  $<1\%$  explained by farming system and by 44, 39, and 66% by site  
253 (Figure 2a). The variation in the proportion of root biomass was by 37, 26, and 20% explained by  
254 farming system and 22, 12, and 46% by site in the three soil layers (Figure 2b). Farming system and  
255 site, respectively, accounted for 15 and 40% of the variation in shoot biomass (Figure 2c) and 28 and  
256 57% of the variation in root-shoot ratios (Figure 2d).

257 *Management and pedoclimatic variables*

258 The outcomes of the two evaluation methods (univariate and multivariate analyses) were largely  
259 in agreement, i.e. explanatory variables with high relative importance were also significantly related to  
260 the respective response variable, with few exceptions. Relative importance metrics and relations of all  
261 variables are shown in Figures 3 and 4 and corresponding p-values in Supplementary table 5. Here, we  
262 focus on concordant results for both evaluation methods.

263 In the top, intermediate, and deep layer, respectively, the investigated management and  
264 pedoclimatic variables explained together 78, 74, and 72% of the variation in root biomass and 68, 51,

265 and 70% of the variation in the proportion of root biomass (Figure 3). In the top layer, root biomass and  
266 the proportion of root biomass were strongest related to weed biomass (positive) and mineral N  
267 fertilization intensity (negative; Figure 3). High importance for root biomass was also assigned to soil  
268 organic C (positive) and for the proportion of root biomass to soil bulk density (negative; Figure 3). In  
269 the intermediate layer, sowing density explained the largest part of the variation in root biomass and its  
270 proportion (positive), while root biomass was additionally strongly related to silt content (negative) and  
271 the proportion of root biomass to mineral N fertilization intensity (positive; Figure 3). In the deep layer,  
272 precipitation had the highest importance for root biomass and a strong positive effect, while the  
273 proportion of root biomass was not significantly related to any variable (Figure 3).

274 The investigated management and pedoclimatic variables explained 53 and 88% of the variation  
275 in shoot biomass and root-shoot ratios, respectively (Figure 4). Available soil P was the only variable  
276 with a significant relation (positive) to shoot biomass with high importance, while large parts of the  
277 variation in root-shoot ratios were explained by mineral N fertilization intensity (negative) and weed  
278 biomass (positive; Figure 4).

## 279 4. Discussion

### 280 4.1. Management effects on root biomass allocation to agricultural soils

281 In this comprehensive on-farm study, we found 40% higher total root biomass under organic  
282 compared to conventional farming. This is to our knowledge the first study highlighting this substantial  
283 farming system effect on root biomass allocation in an on-farm setting characterized by a wide range of  
284 management and pedoclimatic conditions across fields. The results thus allow particularly robust  
285 conclusions on farming system effects on root biomass allocation. Moreover, conventional agriculture  
286 in Switzerland relies to a high degree on cultivation practices that are also typical of organic farming  
287 such as long and diverse crop rotations, inclusion of cover crops, and frequent organic fertilization  
288 (Nitsch and Osterburg, 2005). A comparison of more divergent systems (e.g. mono-cropping with sole  
289 mineral fertilization vs. long crop rotations with sole organic fertilization) might reveal even more  
290 pronounced farming system effects. Hence, the here presented results constitute rather conservative  
291 estimates for enhanced root C allocation through organic farming in agroecosystems.

292 This study therefore provides supportive evidence for higher root C inputs into organic  
293 compared to conventional soils, which has also been found by Chirinda *et al.* (2012) and Hu *et al.* (2018)  
294 at several long-term field sites in Denmark. Those and our findings suggest an effect size of plus 20–  
295 40% root biomass in organic compared to conventional systems and thereby oppose the currently  
296 prevailing view that organic farming reduces root C inputs along with yields (Lorenz and Lal, 2016). In  
297 our study, shoot biomass at flowering showed only a small, non-significant difference among organic  
298 and conventional farming and grain yield at harvest was even about 30% lower on the organic than  
299 conventional fields (Büchi *et al.*, 2019). Consequently, biomass allocation below and above ground  
300 follows different patterns in organic and conventional systems.

301 The farming system effect on total root biomass was mainly a composite of effects of three  
302 management-related factors on root biomass in the topsoil. Among the most important drivers was weed  
303 biomass, which was an order of magnitude higher in organic (56 g m<sup>-2</sup>) than conventional farming (5 g  
304 m<sup>-2</sup>). Weed roots can trigger over-proliferation of crop roots (Depuydt, 2014) when crops and weeds  
305 compete for the same below ground resources (Kiær *et al.*, 2013). However, information on root biomass  
306 of weeds would be inevitable to clearly disentangle physiological and methodological causes. As fibrous  
307 roots of weeds and crops are often not distinguishable by eye, precise classification requires elaborate  
308 methods (Watt *et al.*, 2008; Hirte *et al.*, 2017). As we could remove only clearly identifiable weed roots  
309 from the root samples, we assume that weed roots have partly altered sample weight. As a conservative  
310 estimate from our weed shoot biomass data and published root-shoot ratios of weeds that correspond to  
311 total weed root biomass (Blackshaw *et al.*, 2003; Moreau *et al.*, 2017; Hu *et al.*, 2018), we consider  
312 weed root biomass in the organically managed soils to be at most 25 g m<sup>-2</sup>, thus potentially accounting  
313 for up to 50% of the surplus root biomass in organic compared to conventional farming. The presence  
314 of weeds, however, is an important aspect of management and contributes in real terms to root biomass  
315 and thus organic C inputs to soil.

316 Similarly important for topsoil root biomass was mineral N fertilization intensity, which was  
317 40% lower on the organic than conventional farms. Low mineral N availability in soil has previously  
318 been found as the main reason for higher root biomass in organic compared to conventional farming  
319 (Chirinda *et al.*, 2012; Hu *et al.*, 2018). In mineral N limited systems, crops invest a larger proportion of

320 assimilates in below ground organs in order to increase plant interception of soil-borne resources (Lynch  
321 *et al.*, 2012). By contrast, total soil N was not related to root biomass in our study, indicating that this  
322 variable, unlike mineral N fertilization intensity, did not represent available soil N fractions adequately.  
323 The importance of available soil P for root biomass was similarly low despite its strong positive effect  
324 on shoot biomass. Phosphorus supply influences rooting characteristics predominantly by altering  
325 topsoil root proliferation, whereas root biomass is only affected under severe P shortage (Hermans *et*  
326 *al.*, 2006). This highlights the outstanding role of N nutrition in the studied farming systems.

327         Soil organic C was the third factor that was prominently related to topsoil root biomass.  
328 Although it differed more strongly among sites than farming systems, it was elevated in the organic  
329 compared to the conventional soils. This difference proved to be significant in the extended farm  
330 network which also included the farms from this study (Colombi *et al.*, 2019). Higher soil organic C can  
331 be a consequence of higher root biomass or *vice versa* as the underlying processes can be bi-directional.  
332 On the one hand, continuously increased root biomass enhances soil organic C in the long-term (Lajtha  
333 *et al.*, 2014) due to its strong influence on soil organic matter formation (Rasse *et al.*, 2005; Kätterer *et*  
334 *al.*, 2011; Menichetti *et al.*, 2015). On the other hand, higher soil organic C can improve soil aeration  
335 and thus stimulate root growth (Colombi *et al.*, 2019). Methodological aspects of sample processing can  
336 also entail spurious relationships between soil organic C and root biomass when root samples contain  
337 large amounts of extraneous organic matter due to e.g. frequent organic fertilization (Hirte *et al.*, 2017).  
338 However, as C inputs to soil by crop residues and organic fertilizers were not substantially increased on  
339 the organic compared to the conventional farms (Colombi *et al.*, 2019), we assume a causal relationship  
340 between higher root C inputs and increased organic C content in the organically managed soils.

341         Root biomass in no-till soils was intermediate and not significantly different from that in  
342 conventionally and organically managed soils. Interestingly, it was markedly elevated by data from one  
343 farm (274 g m<sup>-2</sup>) that used a seed mix of two wheat varieties. Knowledge on root traits in mixed wheat  
344 stands is scarce but findings for other crops suggest that competition between genotypes in mixed stands  
345 increases biomass allocation below ground compared to single stands (Ninkovic, 2003; Lin *et al.*, 2014).  
346 As revealed by the medians, root biomass in no-till farming (138 g m<sup>-2</sup>) was actually much closer to that  
347 in conventional (118 g m<sup>-2</sup>) than that in organic farming (178 g m<sup>-2</sup>). This lack of tillage effects on root

348 biomass and, consequently, root-shoot ratios supports previous findings (Anderson, 1988; Williams *et*  
349 *al.*, 2013; Plaza-Bonilla *et al.*, 2014). However, several studies have reported a shift in vertical root  
350 distribution due to no-till (Dwyer *et al.*, 1996; Ball-Coelho *et al.*, 1998; Barzegar *et al.*, 2004), which  
351 we did not observe. Despite a clear relation to soil bulk density in the top layer, the proportion of topsoil  
352 root biomass differed by only 2% between no-till and conventional farming in our study. Instead, weed  
353 biomass and mineral N fertilization intensity were the main drivers of vertical root distribution and  
354 accounted for the increased proportion of topsoil root biomass by 8% in the organically managed soils.

#### 355 4.2. Pedoclimatic drivers of root biomass

356 Management effects on total root biomass resulted solely from the large differences in root  
357 biomass between organic and conventional fields in the topsoil, where farming system explained 32%  
358 of the variation. This decreased to basically zero in the subsoil, reflecting the lack of differences in root  
359 biomass between farming systems below 0.25 m depth. In contrast to farming system, site governed root  
360 biomass not only in the top layer but most prominently in the deep layer, where it accounted for 66 and  
361 46% of the variation in root biomass and the proportion of root biomass, respectively. Although the sites  
362 spread over a distance of just 100 km, their edaphic characteristics varied strongly, representing the  
363 diversity of European soils (Ballabio *et al.*, 2016; Ballabio *et al.*, 2019).

364 Below 0.25 m soil depth, spring precipitation became increasingly important for root biomass  
365 and explained even 40% of its variation in the deep layer. We infer that water was not limiting at any of  
366 the studied fields as rainfall was 150 mm (50%) higher than mean annual precipitation (30-year climate  
367 norm) from April to June 2016. The particularly moist spring conditions even caused below-average  
368 yields (Büchi *et al.*, 2019), which was possibly linked to fewer sunshine hours, higher pest and disease  
369 pressure, and fewer opportunities for farmers to perform mechanical soil cultivation for e.g. weeding.  
370 Instead, since rainfall is one of the most important driving forces of nitrate leaching in agroecosystems  
371 (Goulding *et al.*, 2000; Jabloun *et al.*, 2015), the strong positive relation between precipitation and deep  
372 root biomass could be an indication of root response to relocation of N.

373 Subsoil root biomass was also prominently linked to soil texture, in particular silt content in the  
374 intermediate layer and sand content in the deep layer, which ranged between sites from 29 to 40% and

375 31 to 54%, respectively. Their respective negative and positive effects on subsoil root biomass support  
376 findings of greater rooting depth in coarse- than medium-textured soils in temperate climate (Schenk  
377 and Jackson, 2005). The unfavourable capacity of sandy soils to hold plant-available water and nutrients  
378 forces plants to root deeper in order to meet their demand for those resources. In our study, higher  
379 nutrient availability in silty soils was likely to result in lower investment of wheat in root growth below  
380 the topsoil, which has also been reported from two Swiss long-term field trials (Hirte *et al.*, 2018a).

381 Sowing density, which was the only driver of root biomass entirely independent of farming  
382 system and site, had a strong positive impact in the intermediate soil layer. While it has previously been  
383 shown that root biomass in the topsoil increases with sowing density, no effects have so far been found  
384 in the subsoil (Marcinkevičienė *et al.*, 2013; Hecht *et al.*, 2016). We assume that fertilization and weed  
385 control were the main drivers of root response in the topsoil and overlaid the potential influence of  
386 sowing density on topsoil root biomass in our study. Our results indicate that effects of sowing density  
387 are not confined to topsoils but might easily be masked by concurring drivers, which will need to be  
388 addressed in detail in future research.

389 This on-farm study drew on a clustered design with a range of varying cultivation measures to  
390 reflect standard agricultural practice. Hence, unexplored management practices constitute an additional  
391 source of variation in root biomass, both between and beyond farming systems. For instance, our data  
392 were obtained from 15 wheat genotypes, which differed distinctly among and within farming systems.  
393 Most genotypes cultivated in organic farming, such as the variety “Wiwa”, are long-stalked and thus  
394 superior in weed suppression (Dierauer and Klaiss, 2020), but their rooting patterns have yet to be  
395 investigated in detail. Wheat genotypes can vary by a factor of five in root biomass (Mathew *et al.*,  
396 2019), suggesting that the genotype–environment–management triad that profoundly governs above  
397 ground crop parameters (Hillel and Rosenzweig, 2013; Hatfield and Walthall, 2015), also plays a  
398 significant role in below ground biomass allocation. We therefore argue that a major part of the 30%  
399 variation in root biomass, which remained unexplained in our study, may be assigned to genetic drivers.  
400 Thus, future research employing multidimensional networks with completely crossed designs of  
401 genotype x environment x management can allow to disentangle the complex interactions of farming  
402 system and variety in biomass allocation.



403 4.3. Implications for soil C dynamics, soil C modelling, and climate change mitigation

404 Higher root biomass in organic than conventional topsoils implies considerably larger total  
405 below ground C inputs via root biomass and rhizodeposition. The surplus of roughly 25 g m<sup>-2</sup> wheat root  
406 biomass (excluding weeds) in organic farming can be extrapolated to 25 g m<sup>-2</sup> total below ground C  
407 inputs that are additionally allocated to soil by organic compared to conventional wheat in Swiss  
408 agricultural practice (C concentration in wheat roots: 44%; rhizodeposition-root ratio: 1.3; Hirte *et al.*,  
409 2018a; Hirte *et al.*, 2018b). On top of that, weeds provide an extra source of substantial C inputs to  
410 organically managed soils. This stimulates soil organic matter dynamics profoundly, thereby releasing  
411 plant nutrients, providing energy for soil microbes, and contributing to soil organic matter build-up  
412 (Janzen, 2015; Lorenz and Lal, 2016). Hence, by increased topsoil root C inputs, organic farming fosters  
413 soil chemical, biological, and physical processes that enhance soil quality and sustainability of this  
414 agroecosystem.

415 As a consequence of higher root-shoot ratios in organic farming, the well-established approach  
416 in soil C modelling of deriving root biomass from shoot biomass at harvest and plant C allocation  
417 coefficients usually inferred at flowering (Bolinder *et al.*, 1997) may therefore not be suitable for  
418 different farming systems. This is supported by recent studies reporting only poor agreement between  
419 estimated and actually measured root biomass in organic farming (Taghizadeh-Toosi *et al.*, 2016; Hirte  
420 *et al.*, 2018b; Hu *et al.*, 2018). While it has previously been suggested that the major source of this  
421 mismatch is the higher shoot biomass in conventional than organic systems at harvest (Hirte *et al.*,  
422 2018b; Hu *et al.*, 2018), our findings provide evidence that it is further amplified by management-  
423 induced differences in root biomass at flowering. The current use of plant C allocation coefficients in  
424 soil C modelling therefore needs to be revisited, both with regard to farming systems and plant ontogeny.

425 Among the proposed strategies to mitigate climate change through increased C inputs to  
426 agricultural soils (Smith *et al.*, 2014; Paustian *et al.*, 2016), an increase in deep root C is least susceptible  
427 to rapid reversal and therefore of particular importance for long-term C sequestration (Kell, 2012). This  
428 study provides the first robust data on the potential of agricultural management practices to alter deep  
429 root C inputs in the most prevalent arable farming systems in Europe. We give evidence that  
430 pedoclimatic drivers substantially govern root biomass below 0.5 m depth, where the impact of farming

431 system is close to zero. Yet, more than one-third of the variation in subsoil root biomass remains  
432 unexplained, leaving room for prospects to control crop root C inputs to deep layers. We expect that  
433 insights into genetic diversity will contribute to fill this gap and that multidimensional genotype–  
434 environment–management networks should become a central part of future research on soil C  
435 management.

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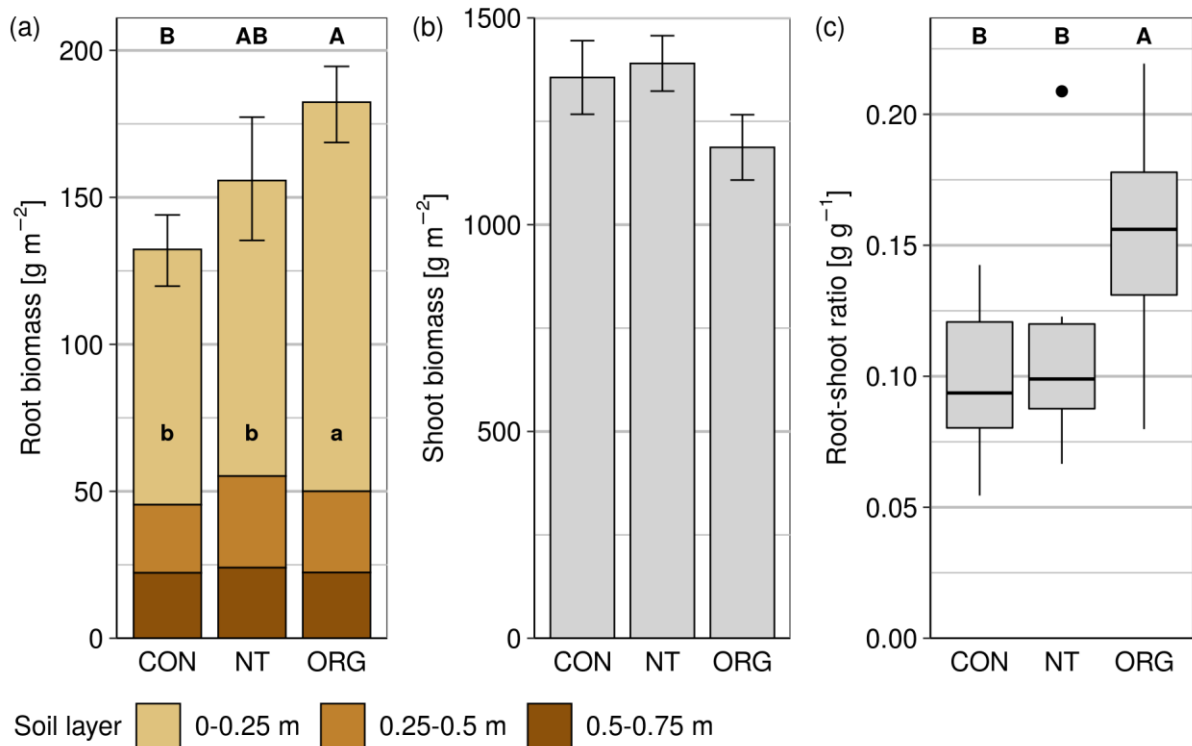
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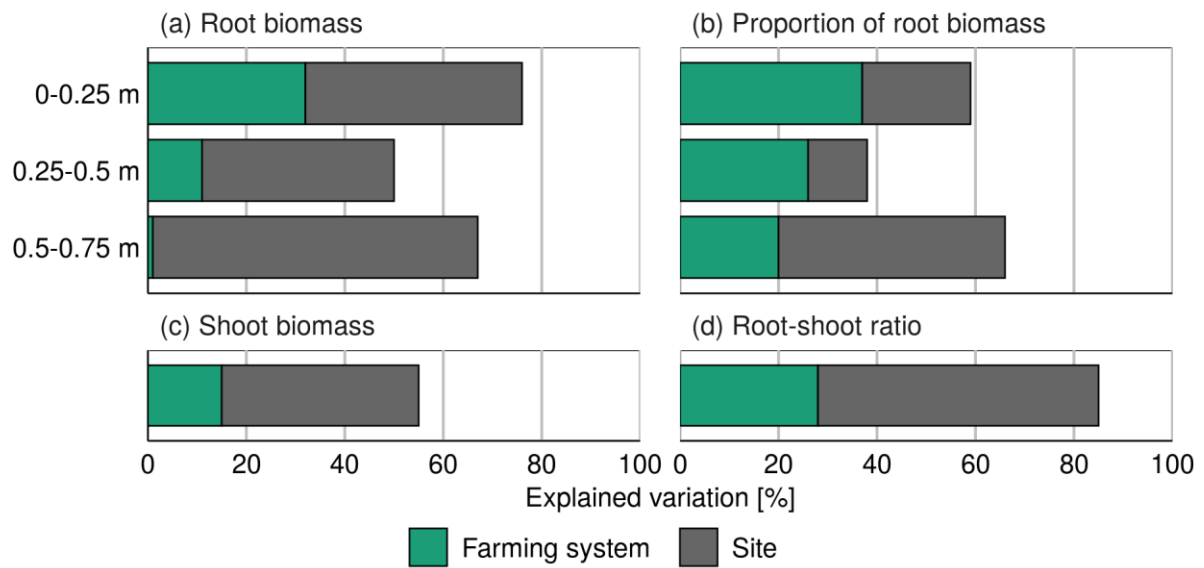
#### 445 **Conflict of interest**

446 The authors declare that they have no known competing financial interests or personal relationships that  
447 could have appeared to influence the work reported in this paper.



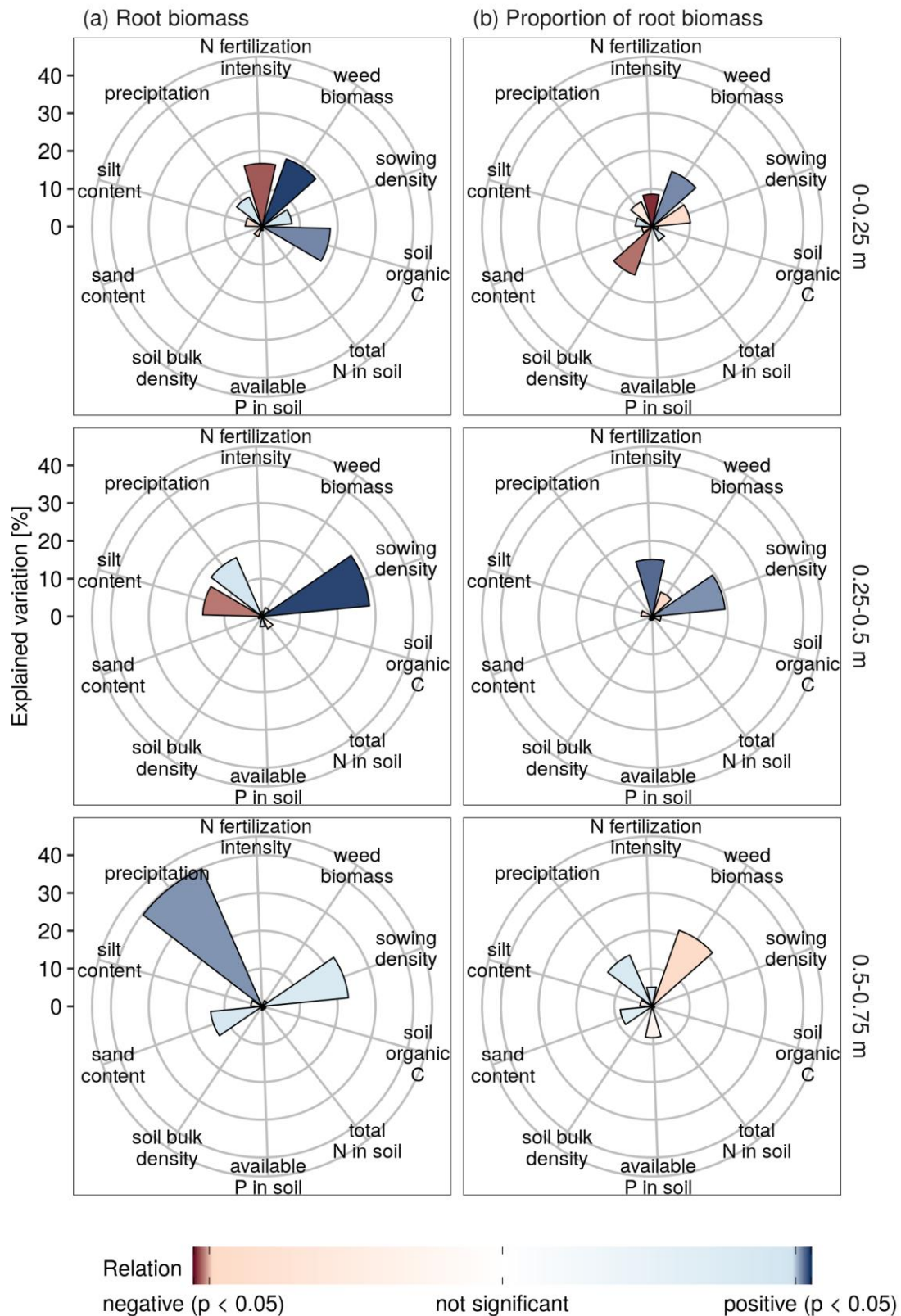
449

450 *Figure 1: Root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–0.75 m) soil*  
 451 *layers, wheat shoot biomass, and root-shoot ratios in conventional (CON), no-till (NT), and organic*  
 452 *(ORG) winter wheat fields at flowering in Switzerland (n = 8 sites; average of 4 field replications each).*  
 453 *Error bars refer to standard errors of total root (0–0.75 m) and shoot biomass of 8 sites. Different letters*  
 454 *denote significant differences between estimated marginal means of root biomass in the individual soil*  
 455 *layers (lower case letters) and total root biomass and root-shoot ratios (upper case letters) at p < 0.05*  
 456 *(Tukey HSD).*



457

458 *Figure 2: Explained variation ( $R^2 * 100$ ) by farming system and site in (a) root biomass and (b) the*  
 459 *proportion of root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–0.75 m) soil*  
 460 *layer, respectively, (c) wheat shoot biomass, and (d) root-shoot ratios in 24 winter wheat fields in*  
 461 *Switzerland.  $R^2$  decomposition method: LMG metrics.*



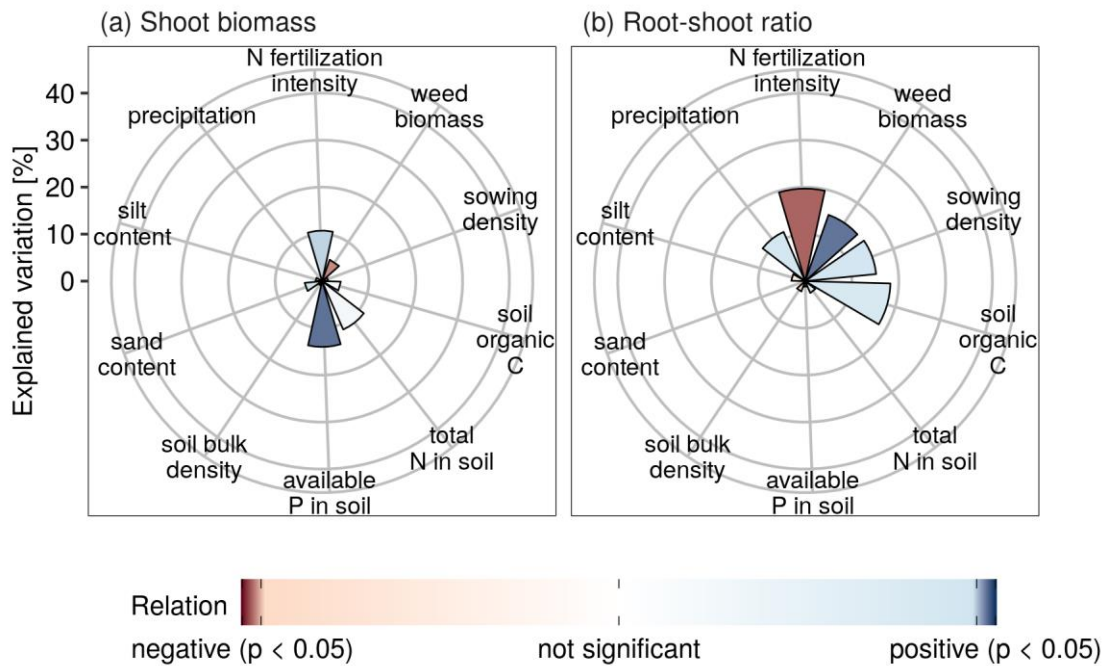
462

463 *Figure 3: Explained variation ( $R^2 * 100$ ) by management and pedoclimatic variables in (a) root biomass*

464 *and (b) the proportion of root biomass in the top (0–0.25 m), intermediate (0.25–0.5 m), and deep (0.5–*

465 *0.75 m) soil layer, respectively, in 24 winter wheat fields in Switzerland.  $R^2$  decomposition method: CAR*

466 scores. Negative / positive relations refer to univariate relations between each management and  
 467 pedoclimatic variable and root biomass (see Supplementary table 5 for p-values).



468

469 *Figure 4: Explained variation ( $R^2 * 100$ ) by management and pedoclimatic variables in (a) shoot*  
 470 *biomass and (b) root-shoot ratios in 24 winter wheat fields in Switzerland (soil variables: top layer).  $R^2$*   
 471 *decomposition method: CAR scores. Negative / positive relations refer to univariate relations between*  
 472 *each management and pedoclimatic variable and shoot biomass or root-shoot ratio (see Supplementary*  
 473 *table 5 for p-values).*

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