

Antimicrobial polymer-clay composite films

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1. Introduction - A principal transmission route for hospital-acquired (nosocomial) infection is via contaminated intermediate vehicles such as equipment and medical devices [1]. Protective antimicrobial polymer coatings can provide some defence against this mode of communication of common hospital pathogens, such as *S. aureus*, *E. coli* and *P. aeruginosa*. Ag⁺-, Cu²⁺- and Zn²⁺-exchanged mineral phases have been used in polymer composites to confer antimicrobial properties on a range of technical and biomedical materials [2]; however, to date, the incorporation of Ag⁺-, Cu²⁺- or Zn²⁺-bearing smectite clay in antibacterial chitosan composites has not yet been investigated. Chitosan affords many advantages over petroleum-based polymers as it is readily abundant, renewable, biodegradable and non-toxic [2]. Accordingly, this research concerns a suite of antimicrobial chitosan composite films incorporating Ag⁺-, Cu²⁺- or Zn²⁺-bearing smectite clay, with potential applications in the control of nosocomial infection.

2. Experimental - Labile Na⁺ and Ca²⁺ ions of natural smectite clay (VEEGUM® F [3]) were exchanged for Ag⁺, Cu²⁺ or Zn²⁺ ions by contacting 1 g of clay with 200 cm³ of 5 mM metal nitrate solutions for up to 48 h at 25 °C [2]. The liquor and solids were separated by filtration and the recovered solutions were analysed by inductively coupled plasma spectroscopy (ICP). As-received smectite or Ag⁺-, Cu²⁺- or Zn²⁺-bearing smectite and chitosan were blended at 5, 10 and 20 wt% in 1% aqueous acetic acid solution (to produce composite films - CS, CSAg, CSCu and CSZn). The films were characterised by scanning electron microscopy (SEM) with energy dispersive X-ray analysis (EDX). The solutions were cast on to polycarbonate surfaces and dried in air at 40 °C. Nutrient agar plates were spread with 0.2 cm³ of *E. coli* (10⁹ CFUcm⁻³). In triplicate, 8 mm discs of each composite were placed in the centre of each plate and the samples were incubated at 37 °C for 24 hours, after which time the zone of inhibition was measured.

3. Results and Discussion - Maximum uptake values of Ag⁺, Cu²⁺ and Zn²⁺ by the clay were found to be 0.72 ± 0.03, 1.90 ± 0.05 and 1.49 ± 0.06 mmol g⁻¹, respectively. Equilibrium was achieved within 6 h. SEM and EDX analysis confirmed that the smectite was uniformly distributed throughout the chitosan matrix and that the films had a fibrous texture (Image 1). Zone of inhibition data are listed in Table I and indicate that composites blended with as-received smectite failed to demonstrate antimicrobial activity against *E. coli*. Conversely, composites containing the ion-exchanged clay showed clear zones, with CSAg and CSZn showing greater inhibitory behaviour than CSCu.

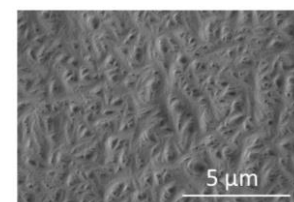


Image 1. SEM image of sample CS with 10% clay-content

Table I. Zone of inhibition data for chitosan-smectite composite films

Sample	CS			CSAg			CSCu			CSZn		
Clay-content	5%	10%	20%	5%	10%	20%	5%	10%	20%	5%	10%	20%
Zone (mm)	0	0	0	0.10 ±0.05	0.17 ±0.06	0.33 ±0.15	0	0	0.100 ±0.001	0.100 ±0.001	0.33 ±0.06	0.33 ±0.06

4. Conclusions - Antibacterial films can be prepared by solvent casting mixtures of chitosan and Ag⁺-, Cu²⁺- or Zn²⁺-exchanged smectite. Ag⁺ and Zn²⁺ were superior to Cu²⁺ at inhibiting the growth of *E. coli*.

5. References

- [1] F.R. Crijns, M.M. Keinänen-Toivola and C.P. Dunne, *J. Hosp. Infect.*, **95**(3), (2017) p. 243.
- [2] A.P. Hurt, A.K. Kotha, V. Trivedi and N.J. Coleman, *Polimeros*, **25**, (2015) p. 311.
- [3] V. Trivedi, U. Nandi, M. Maniruzzaman and N.J. Coleman, *Drug Deliv. Transl. Res.*, **8**(6), (2018) p. 1781.