

Rhomboids and proteolysis in the Dicty mitochondrion

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Rhomboids

Membrane located proteases

- 'Unusual'

Ubiquitous across evolution

- 'One' in prokaryotes: multigene families in eukaryotes

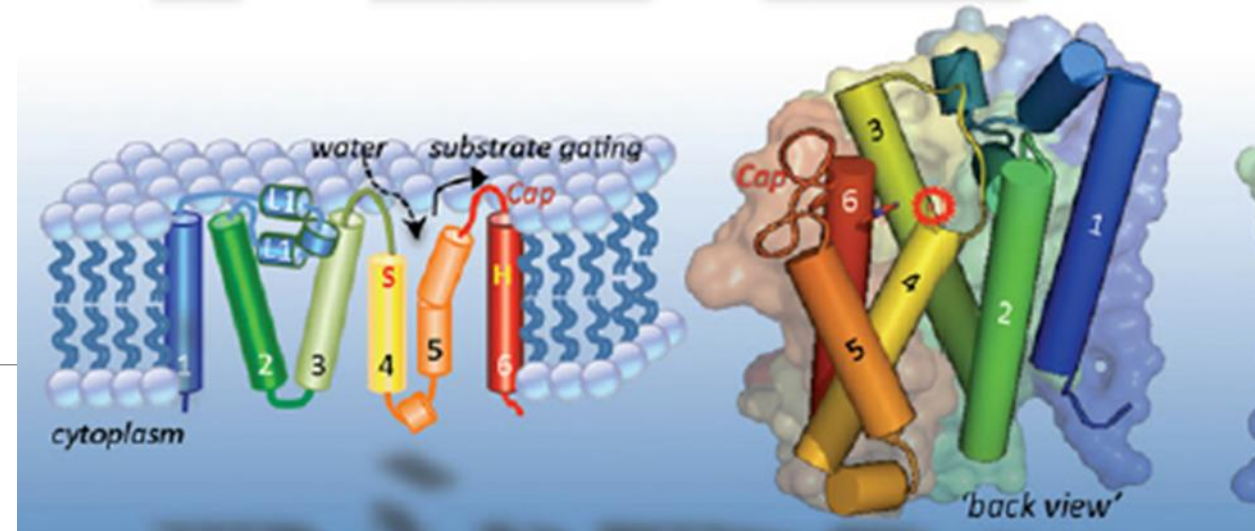
Well-conserved structure

- 6 or 7 t/m; catalytic dyad ...but poor sequence conservation
 - Necessity of conserved motifs not yet fully understood but sequence and structure predicts mitochondrial location of a subfamily
 - And functional vs non-enzymatically active subtypes

Influence development, signalling and infection in a range of eukaryotes and prokaryotes

- e.g., necessary for *Plasmodium* and *Toxoplasma* infection, EGF signalling in *Drosophila*

Substrate prediction is hard



Rhomboid philosophy – why are they interesting?

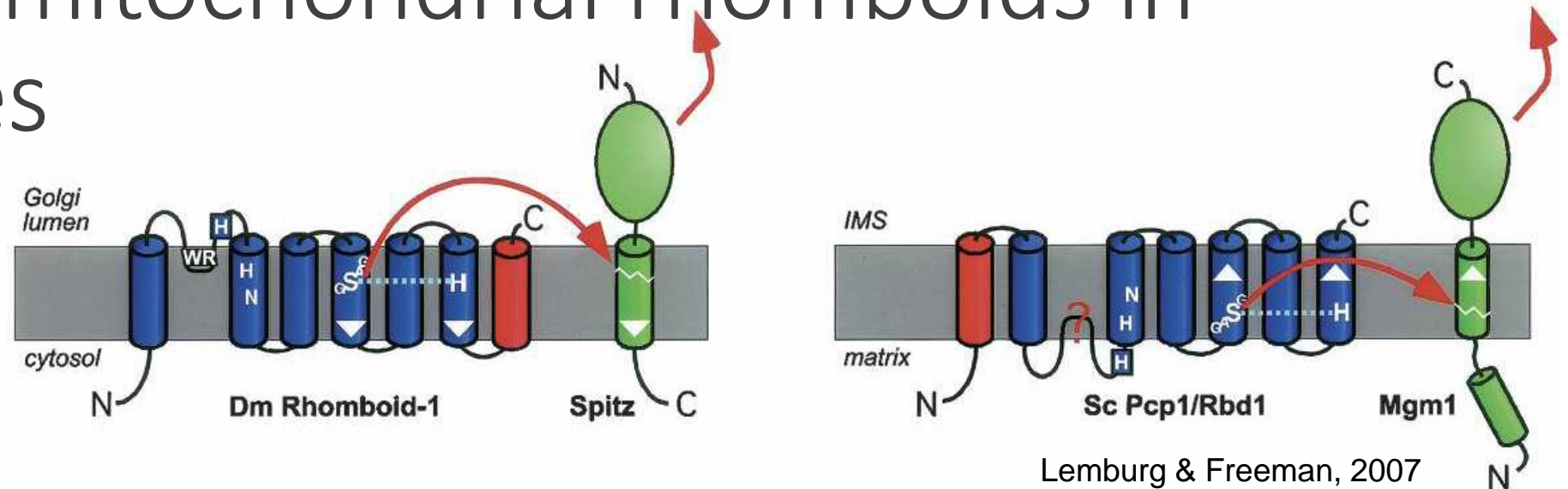
Proteolysis is not just housekeeping: a key regulatory mechanism in cell biology

- Proteases comprise 2-5% of organism genomes across evolution
- Protein activation, localisation, exposure of cryptic binding sites and release of neoproteins...
- Pathogenesis of disease: altered protease expression and substrate-proteolysis, e.g., in Parkinson's and other neurodegenerative diseases

Proteolysis **in cell membranes** occurs via several families of 'new' intermembrane proteases:

- Regulated Interamembrane Proteolysis
- Includes the rhomboid family

Subset of mitochondrial rhomboids in eukaryotes



In *Drosophila* EGFR ligand (e.g., Spitz) cleaved and to release product outside cell for signalling

Orientation reversed in mitochondrial rhomboids

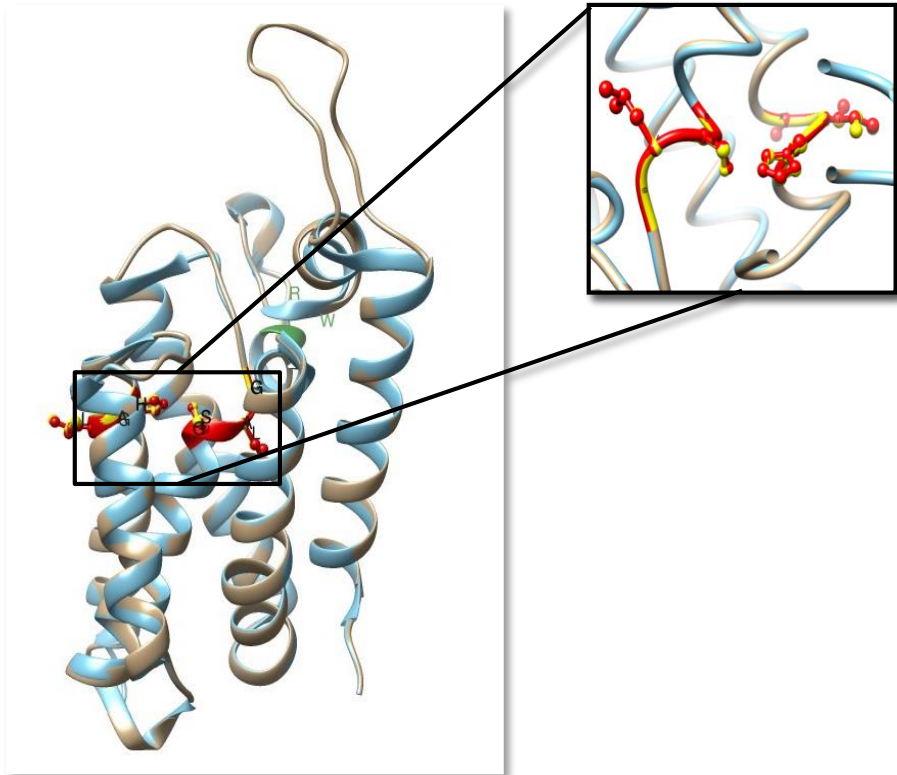
- Rhomboid cleavage is on opposite side of membrane
- Prokaryotes?

Rhomboid subsets in *Dictyostelium*

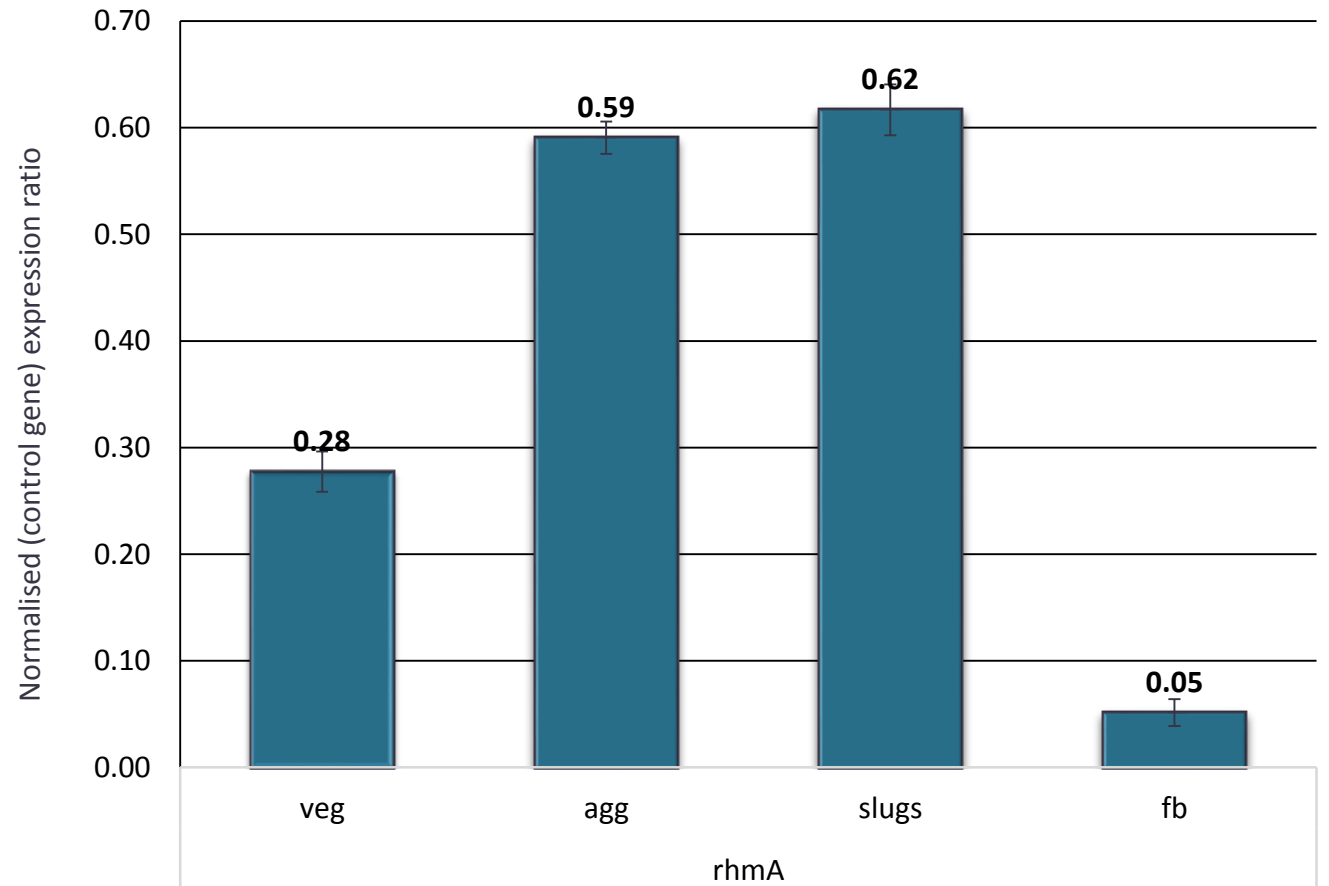
Predicted *Dictyostelium* active rhomboids

| | Sequence ID | |
|-----------|-------------|---------------------|
| Rho like | rhmA | <u>DDB_G0295849</u> |
| Mito like | rhmB | <u>DDB_G0284937</u> |
| Rho like | rhmC | <u>DDB_G0281359</u> |
| Mito like | rhmD | <u>DDB_G0292430</u> |

rhmA – ‘active’, transcription peaks from unicellular-slug stages

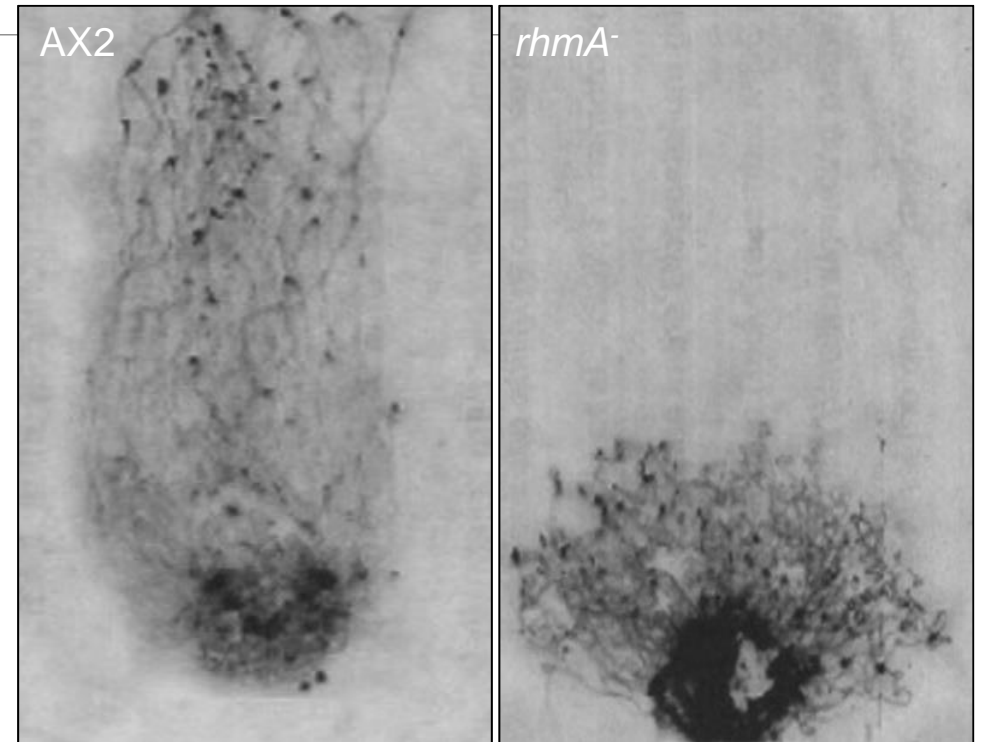


RhmA (brown) vs. *E. coli* GlpG (2IC8, blue). WR (green); G*SG (red)

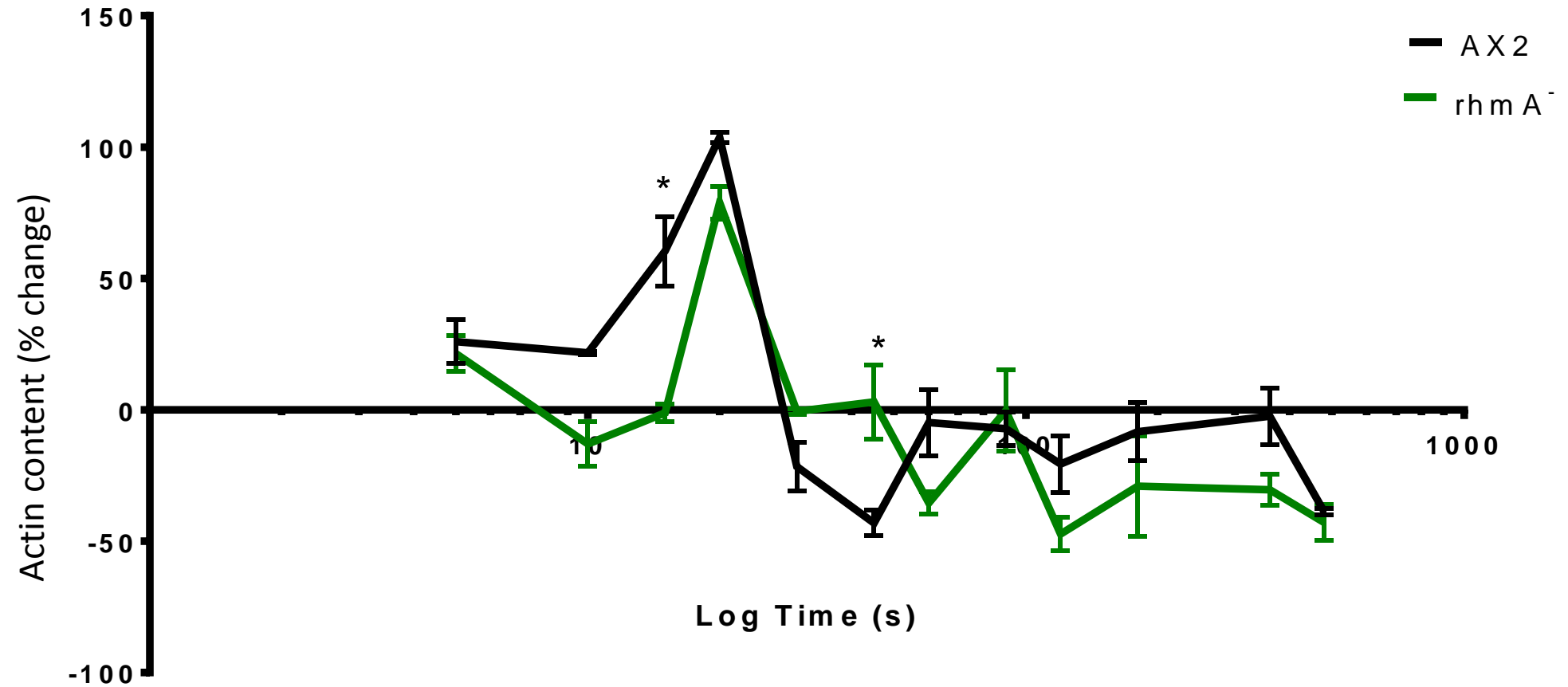


RhmA- phenotype

- aberrant phototaxis,
- slower chemotaxis to both cAMP and folate,
- significant although small reduction in directed movement (random cell movement the same)

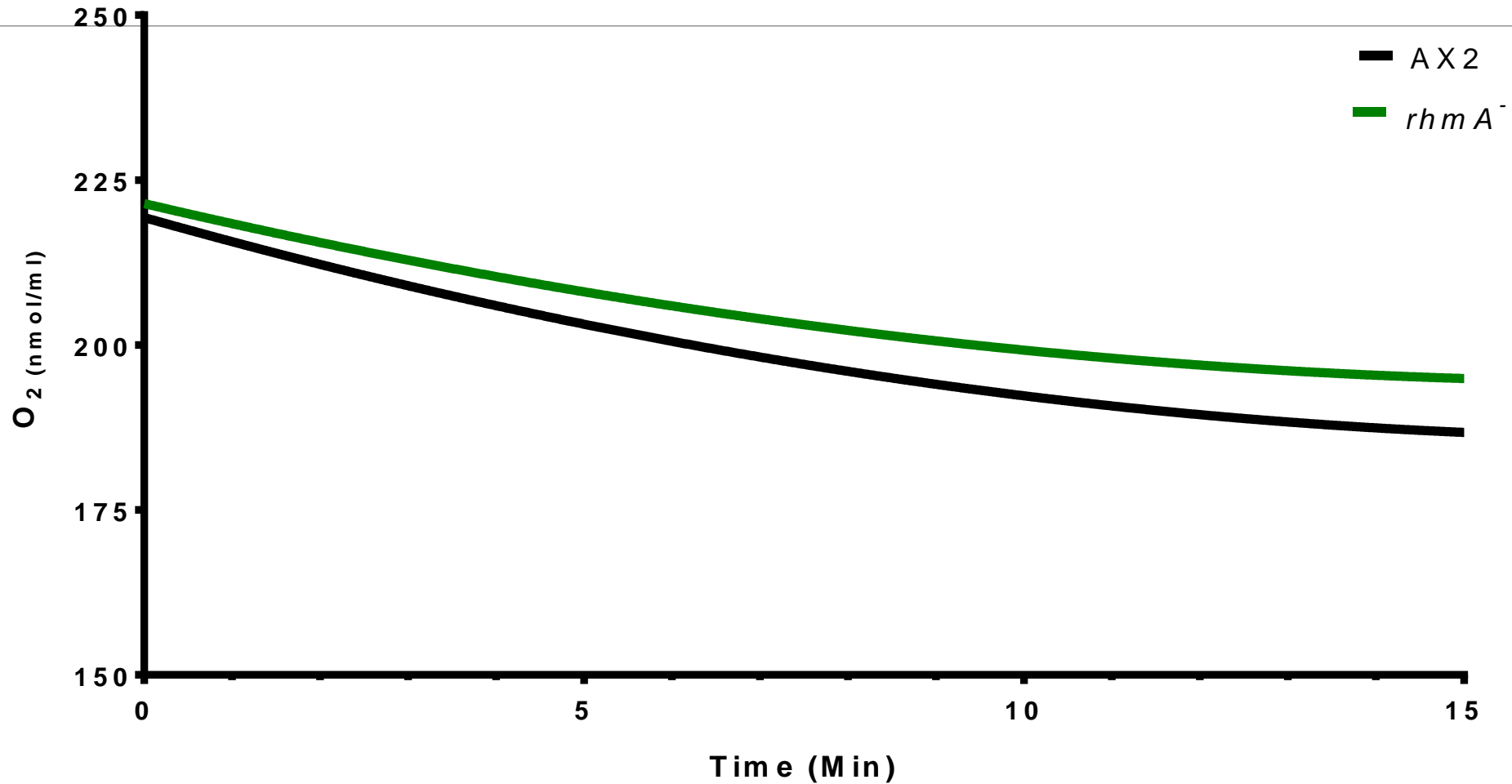


Why defective motility – no actin defect

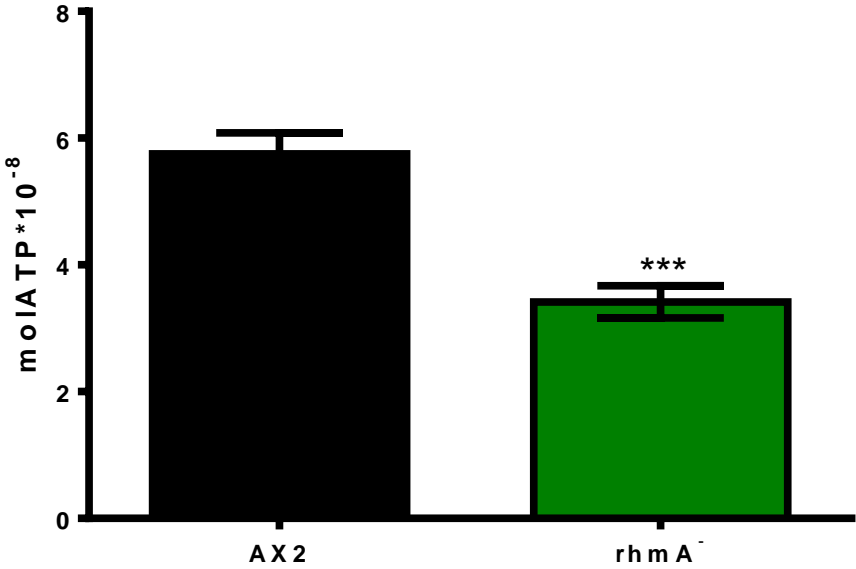


Respiratory defect? Not apparent via O₂ uptake

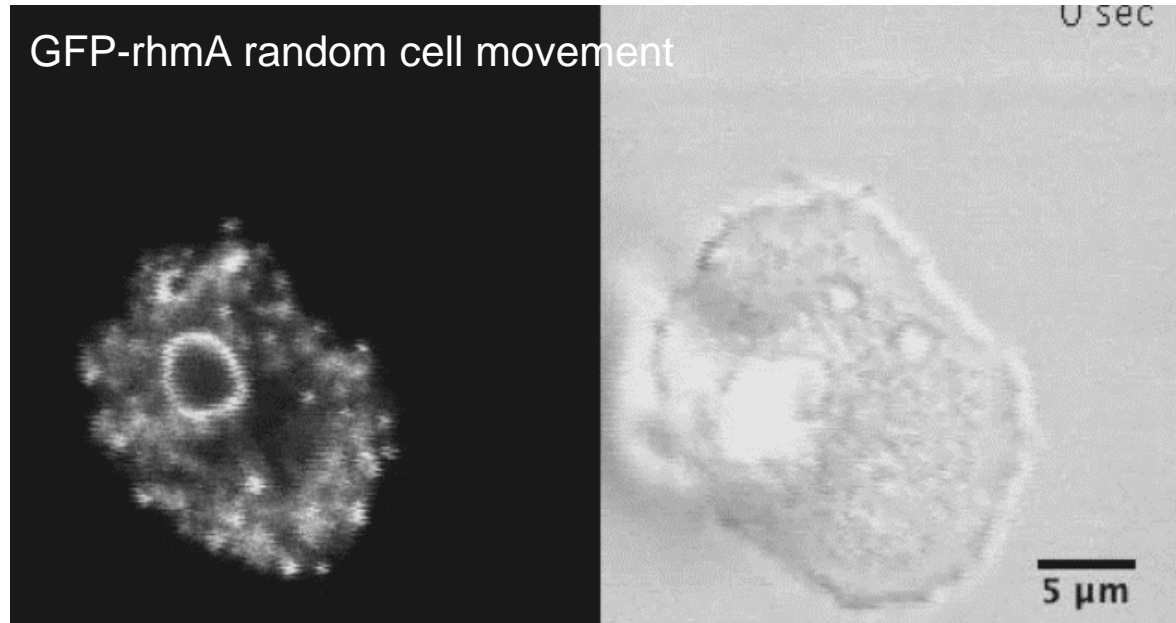
Oxygen Consumption



ATP luciferase assay

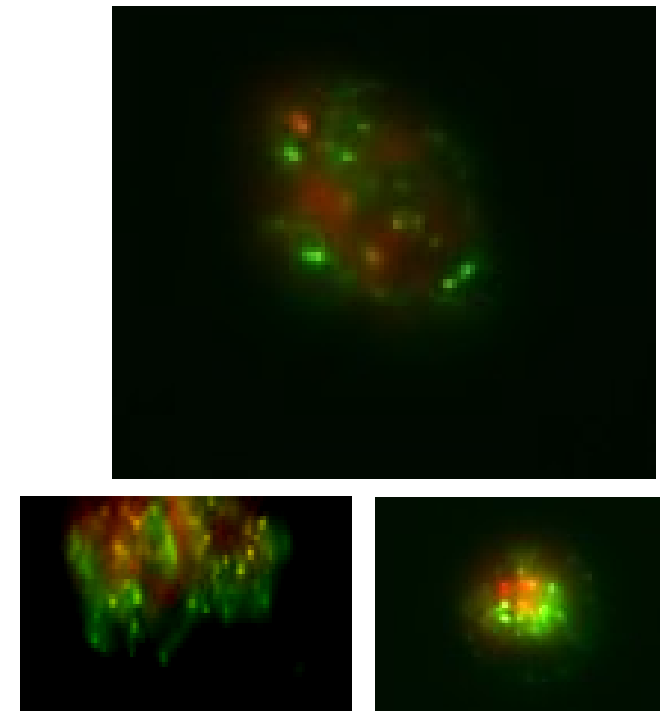


Where is RhmA? GFP-RhmA in contractile vacuole/ cytoplasmic spots



No co-localisation with Mitotracker red

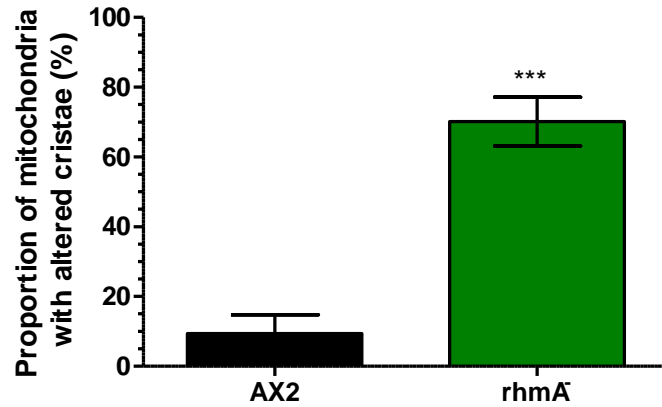
Reduced Mitotracker uptake in rhmA- (MMP?)



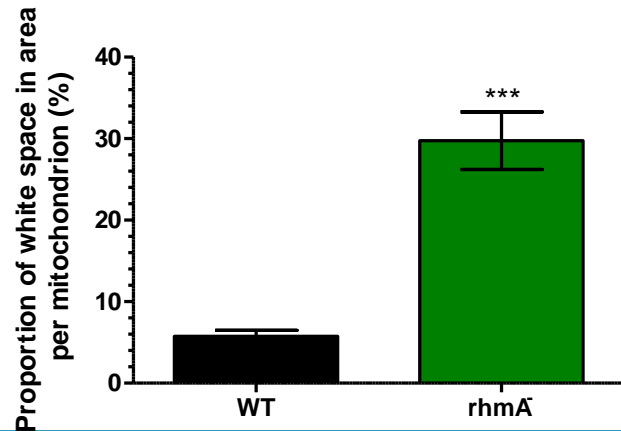
GFP-rhmA (green) co visualised with Mitotracker red

rhmA- mitochondrial morphology defect

A

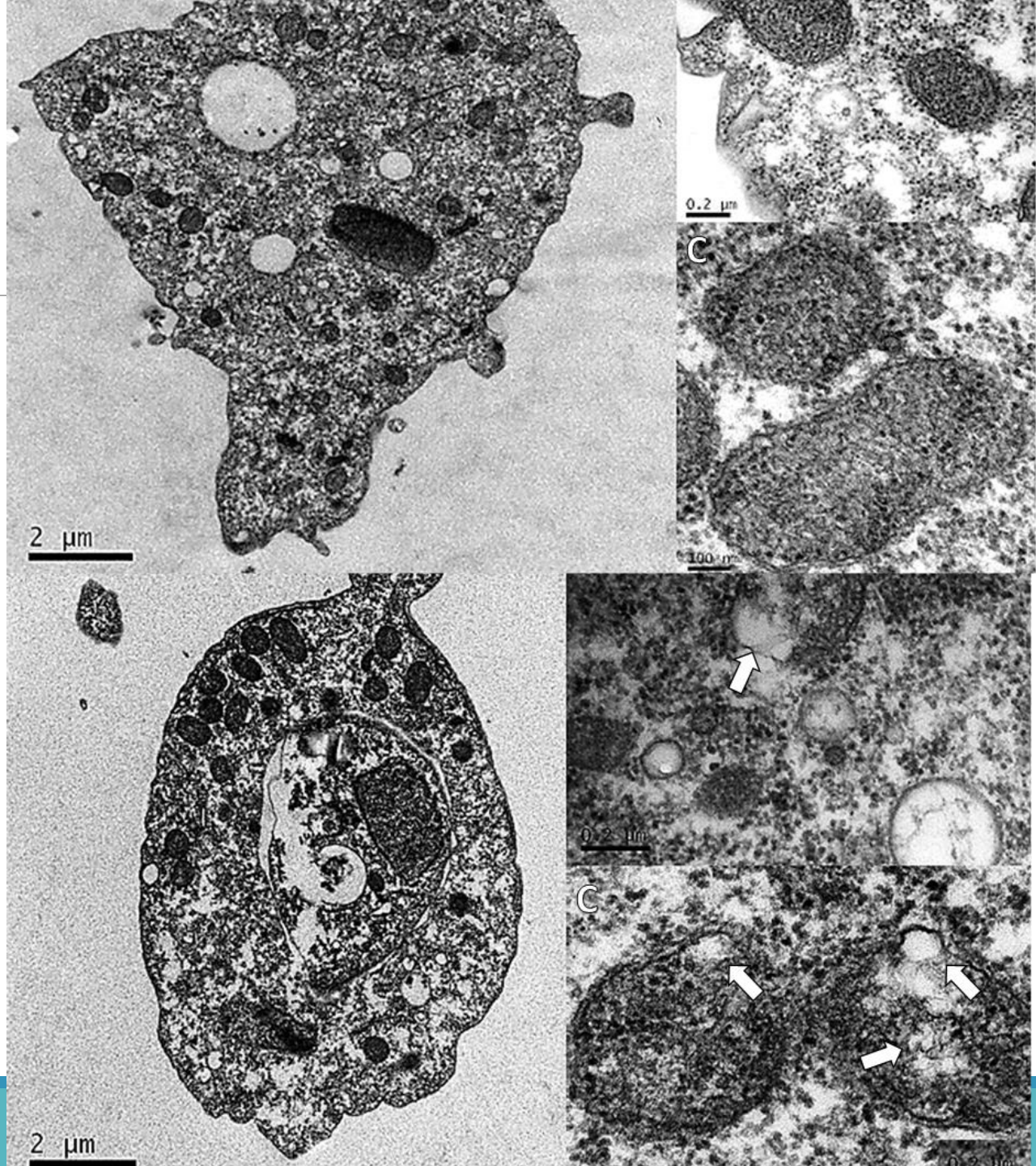


B



Most rhmA- mito show same ultrastructural abnormality

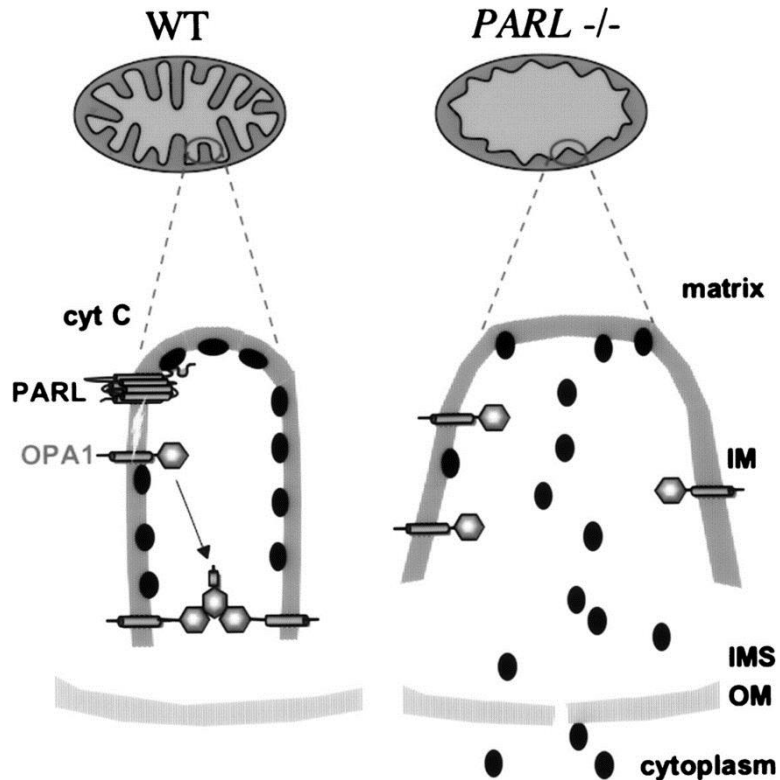
RhmA not predicted/located in the mitochondrion



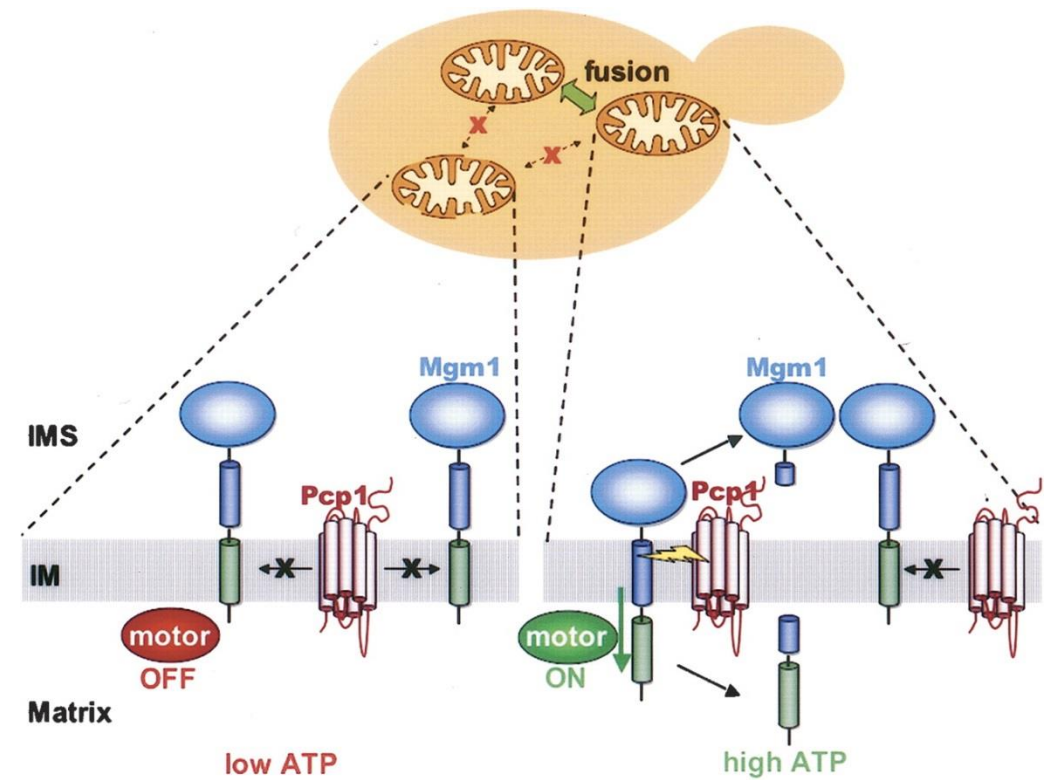
Rhomboid function in mitochondria – dynamin family GTPase substrates

MAINTAINING TIGHT MITOCHONDRIAL CRISTAE IN VERTEBRATES

PARL RBD- CELLS RELEASE CYTC AND HAVE MORE OPEN CRISTAE



REGULATING MITOCHONDRIAL FUSION DYNAMICS IN YEAST



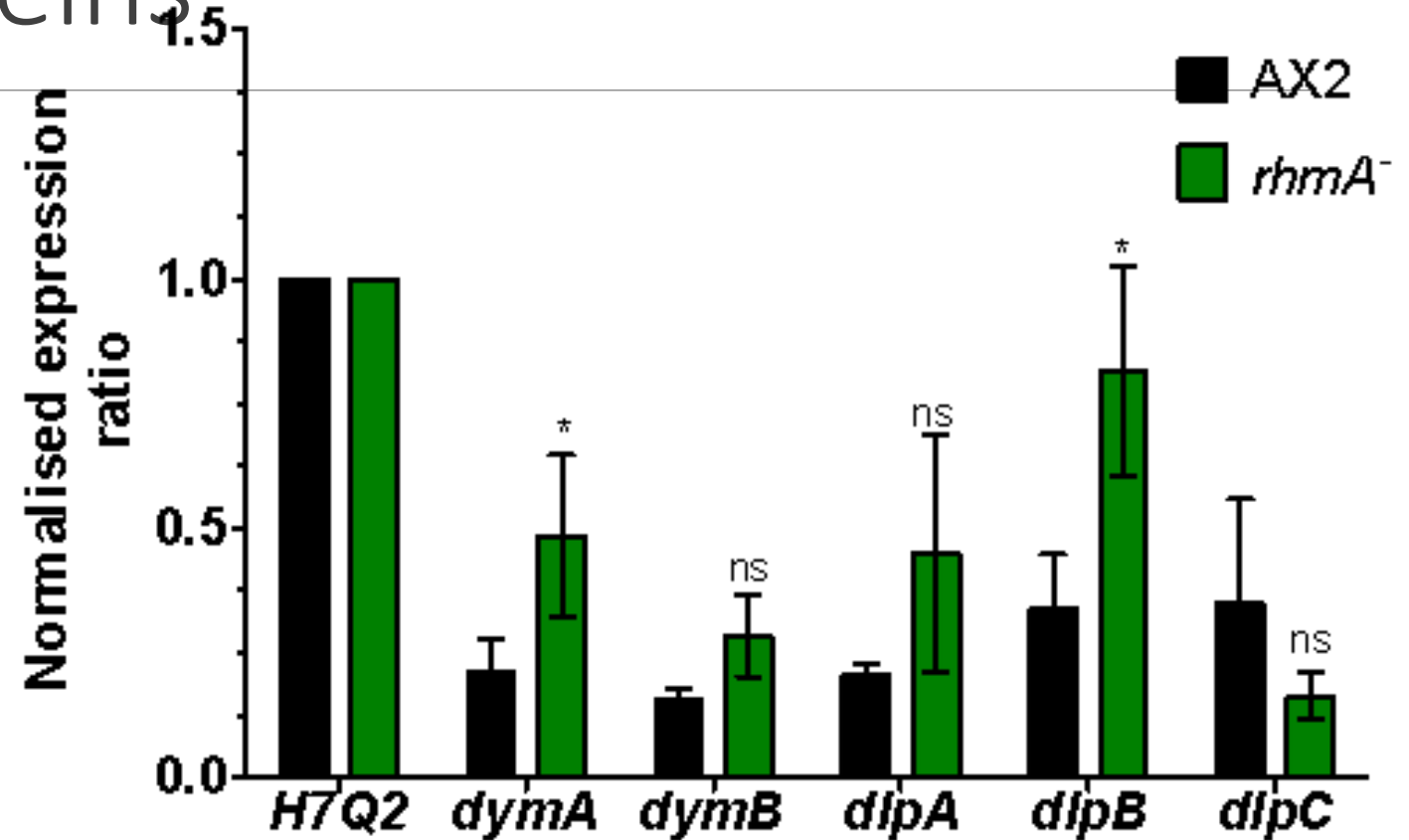
Veg cell RTPCR (no diff in agg cells) -dynamamin-like proteins

Meanwhile...

Pulldown in RhmA-GFP yields band

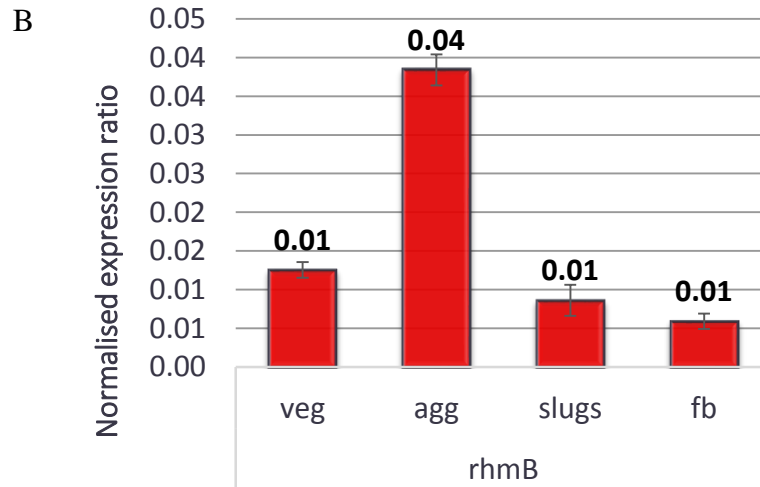
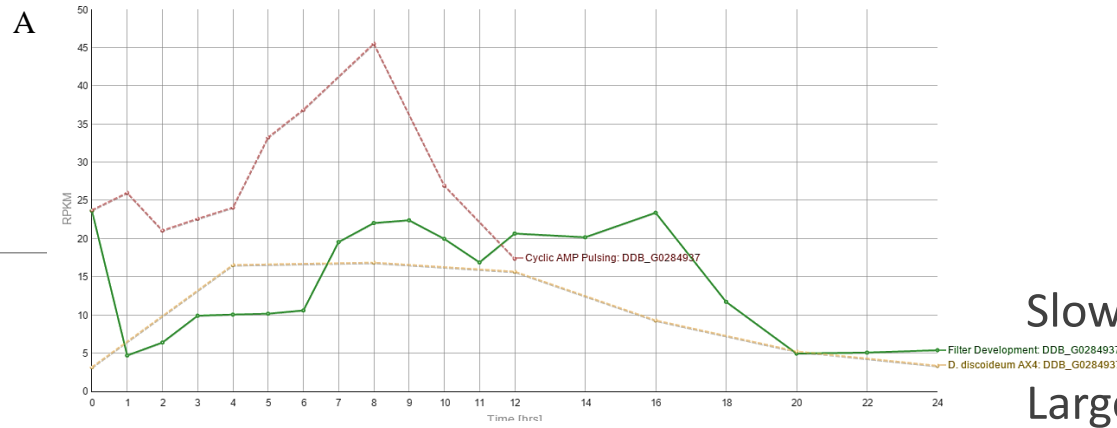
Some ideas...

Activity assay ongoing in Prague



Transcription levels of dynamamin related genes in vegetative cells.

RhmB... succinctly



Slower growth axenic and reduced phagocytosis

Larger cells

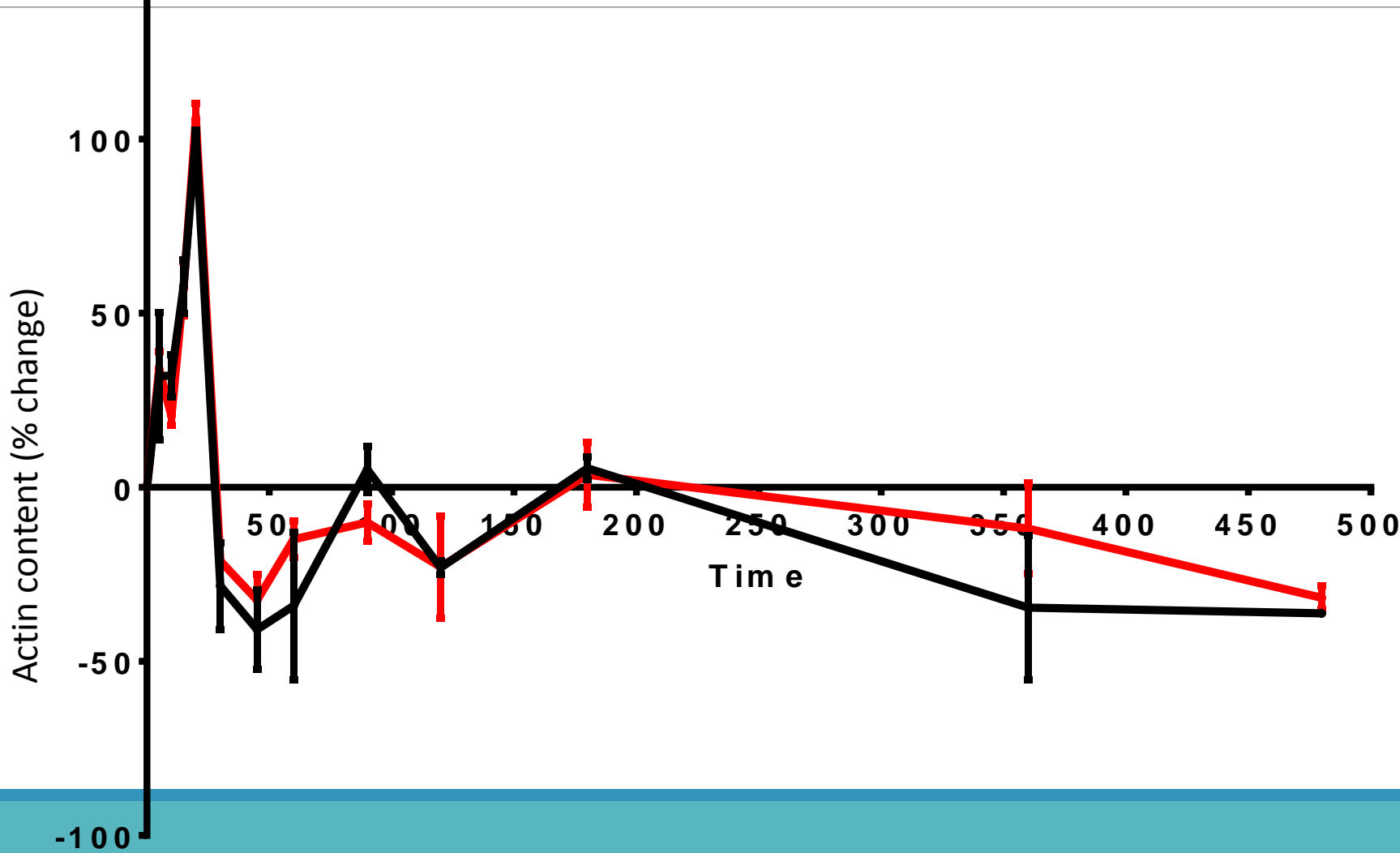
Slower response to folate in one-drop and under-agarose assays

Reduced adhesion – unicellular stage

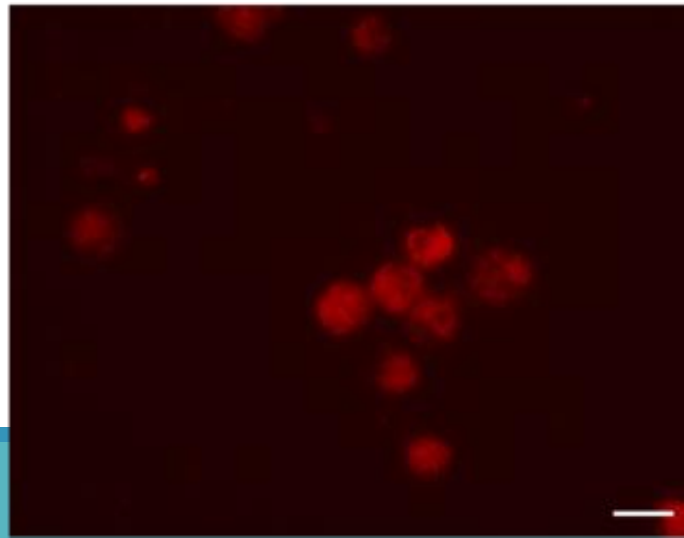
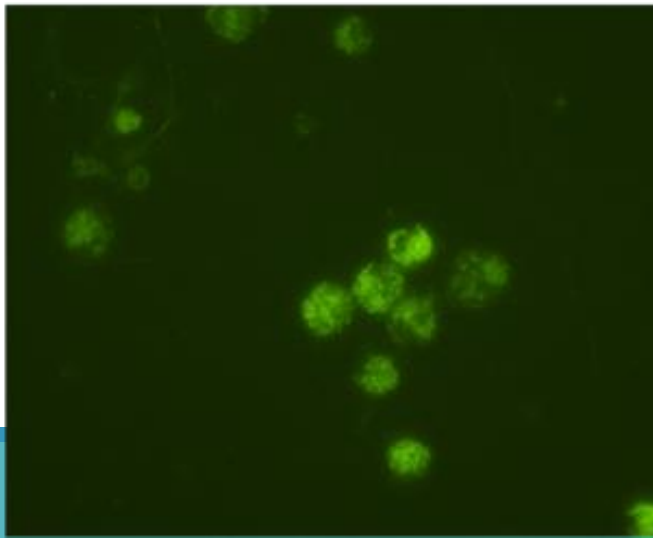
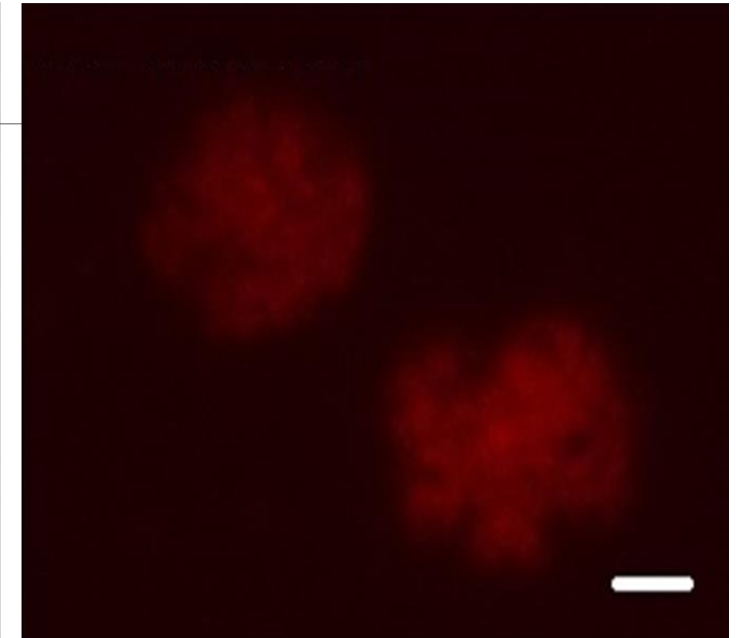
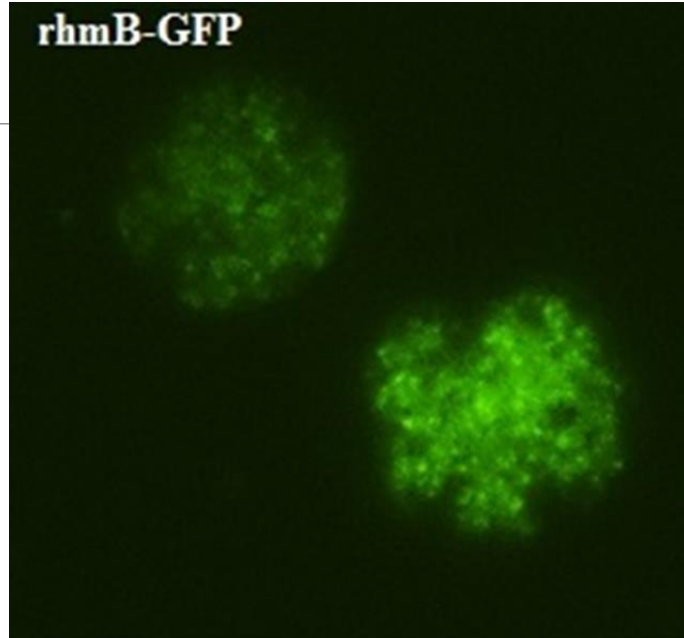
Transcription peak at aggregation (as Dictybase)

Phototaxis as WT

Motility via actin assay

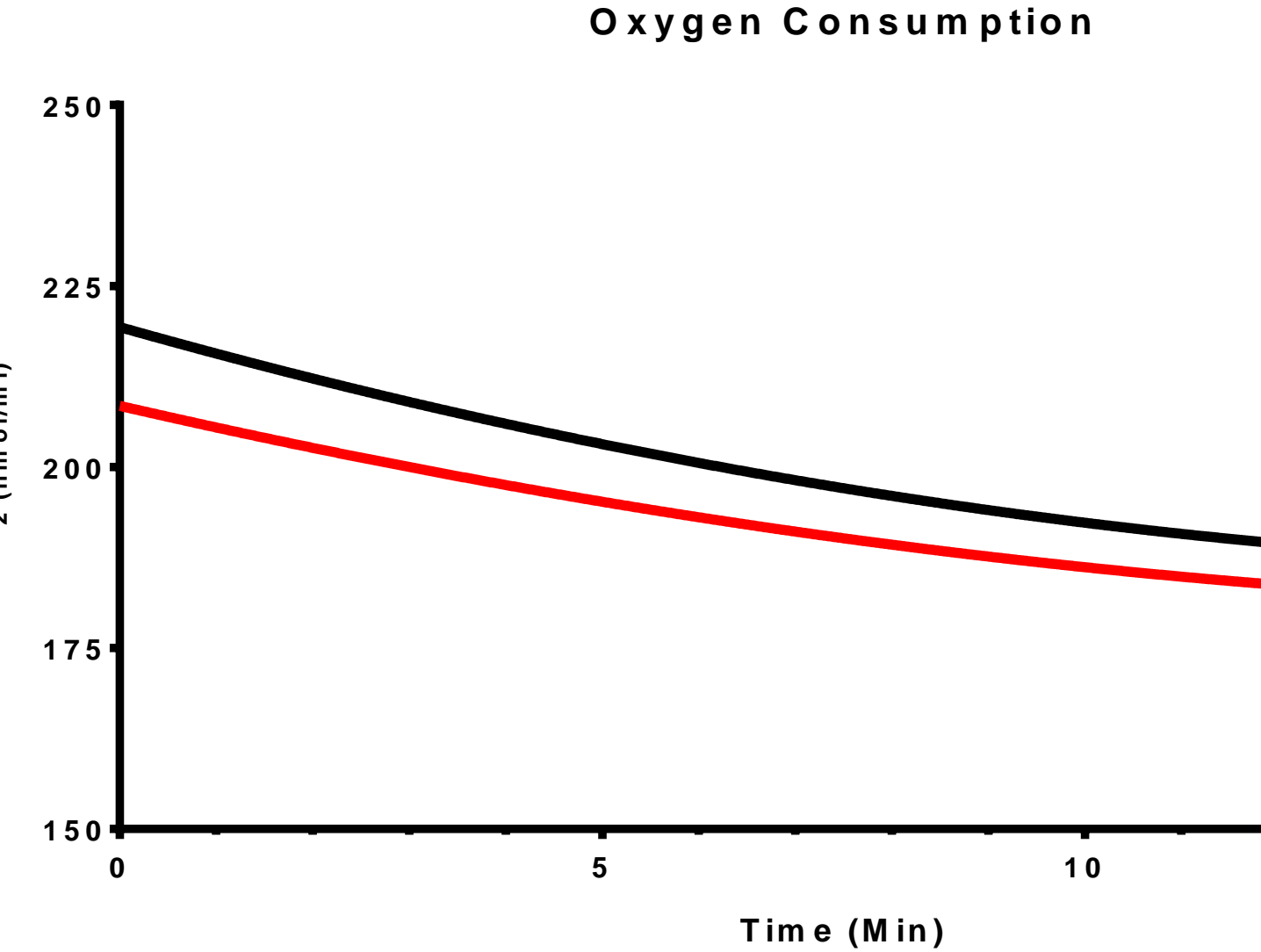


RhmB-GFP fusion protein located in mitochondria ?OM



pJSK543 RFP-GemA mito outer membrane marker

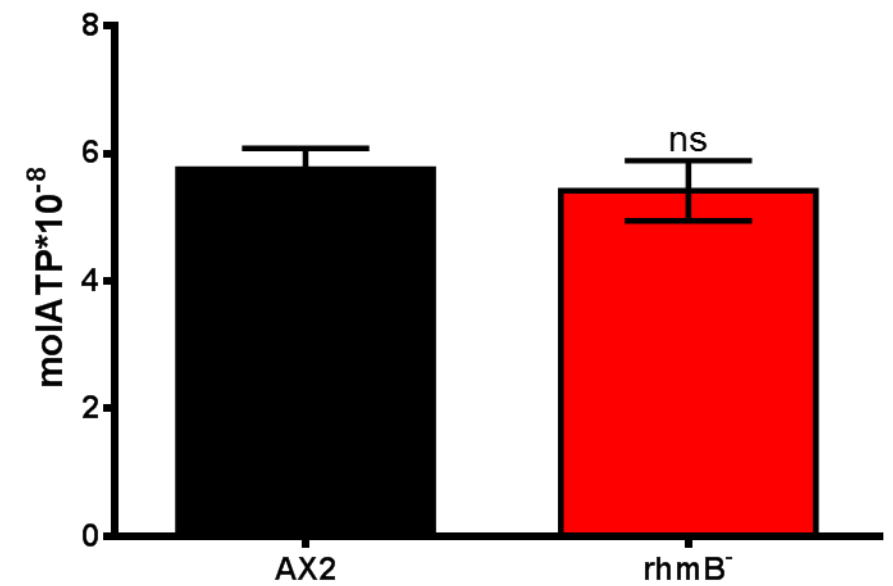
Oxygen Consumption



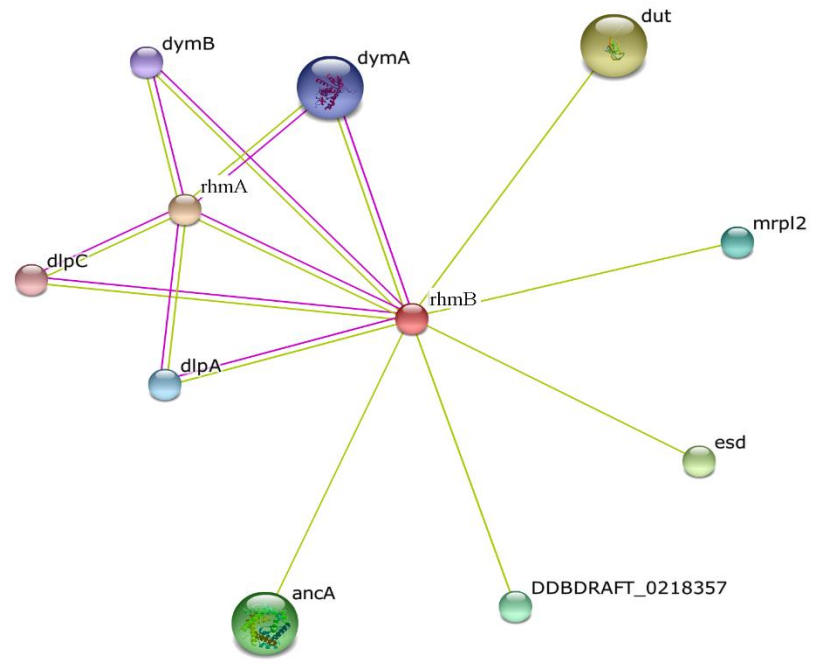
AX2

rhmB⁻

Amount of Atp measured in bioluminescence assay



A



Substrate fishing...

RTPCR with dynamin-related proteins
 Substrate and activity assays ongoing

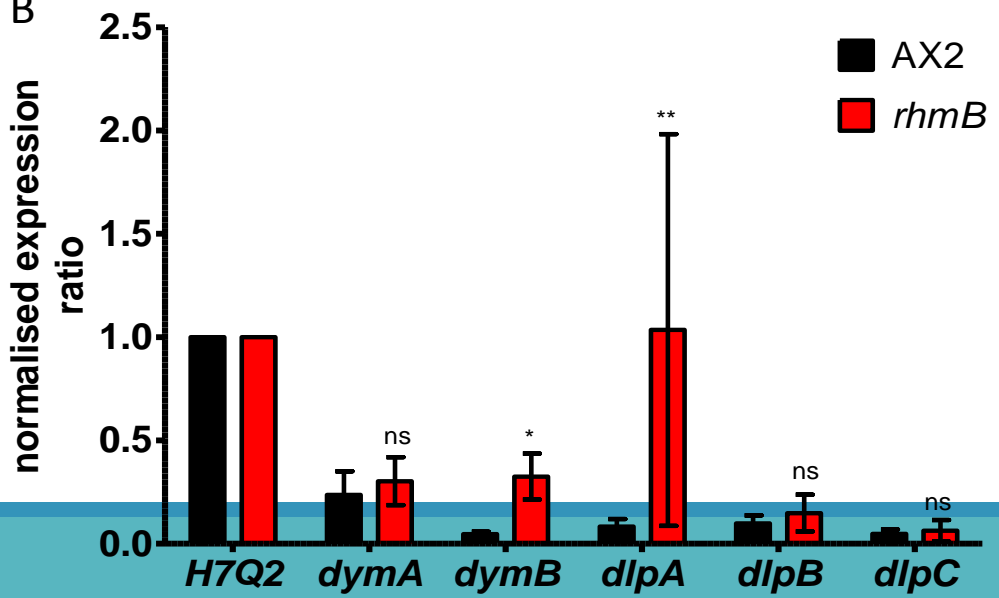
RhmA/B double mutant

No growth on bacterial lawns

RhmD essential

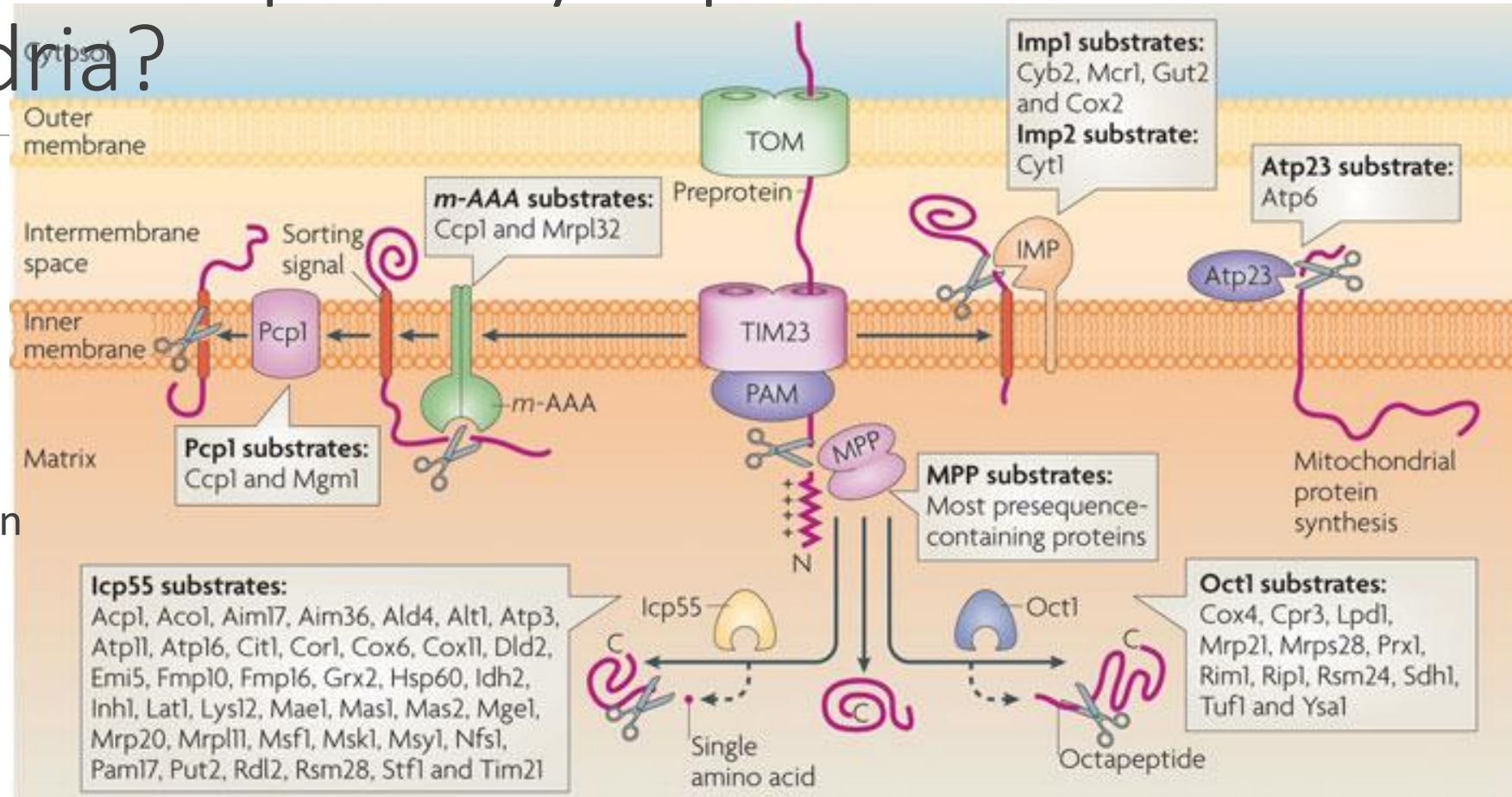
RhmC no pheno?

B



Add rhomboids to the proteolytic proteome of Dicty mitochondria?

- Rhomboids having regulatory roles in *Dictyostelium* mitochondria fits in with our increasing appreciation of the importance of proteolysis in signalling and development – not just house keeping function

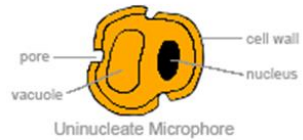


A tendency for specialisation?

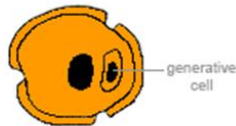
~15 *Arabidopsis rhomboids*

KOM

Microgametogenesis
(Pollen Development)



Uninucleate Microphore



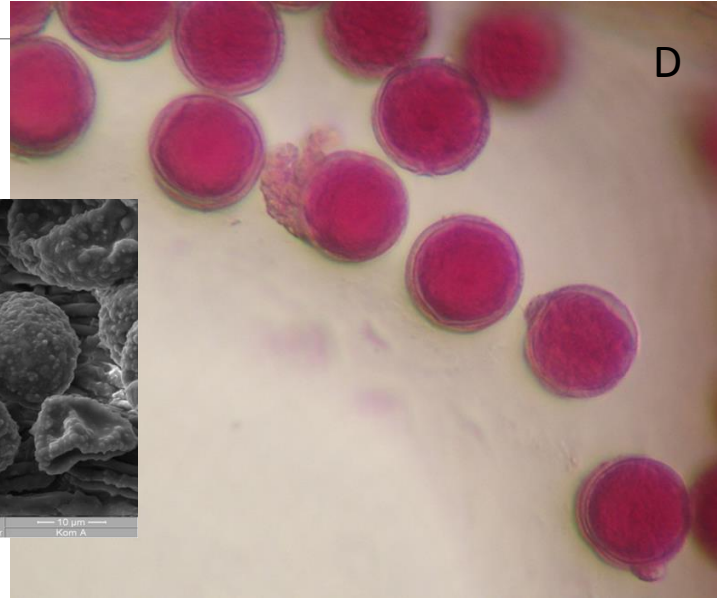
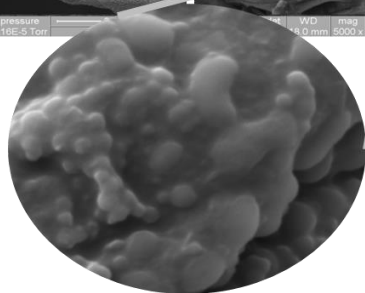
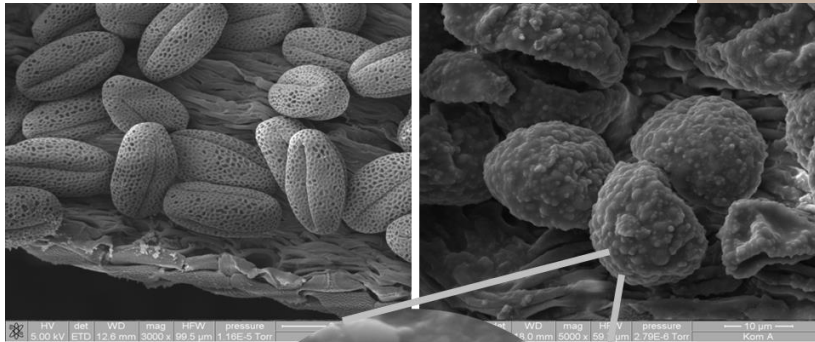
Bicellular Pollen



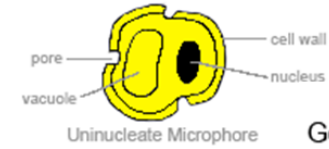
Tricellular Pollen



Mature Pollen Grain



Microgametogenesis
(Pollen Development)



Uninucleate Microphore



Bicellular Pollen



Tricellular Pollen



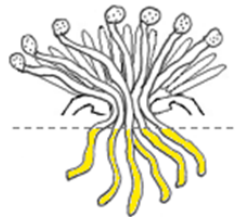
Pollen Germination



30 minutes *in vitro* incubation



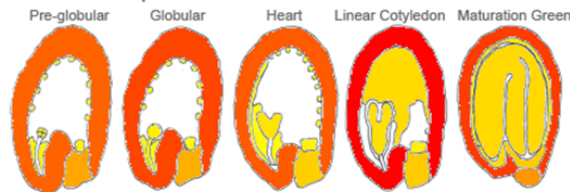
4 hours *in vitro* incubation



Pollen tubes after growth through Stigma and Style (pistil explants)

- Dry pollen grains were harvested by vacuum and germinated using an *in vitro* system, samples were taken at 30 minutes or 4 hours. Pollen tubes were also harvested after germination in a semi-*in vivo* pistil explant system. Samples were taken in quadruplicate or triplicate, and a fluorescence signal is shown. The pollen tube is isolated and hybridized to the ATH1 chip immobilized by GCOS/MAS5, TGT 100.

Seed Development



D



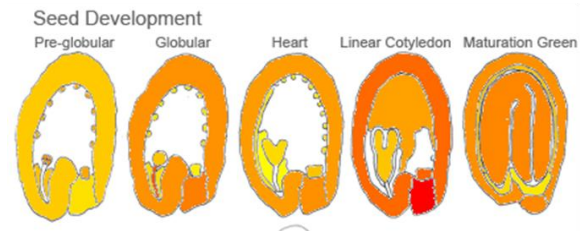
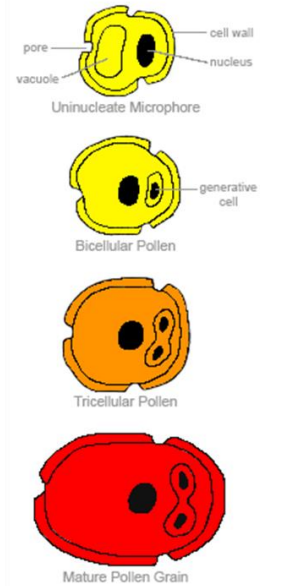
- Developing seeds from *Ws* plants were isolated from continuous light-grown plants and the indicated parts were excised by laser-capture microdissection. - RNA was extracted and amplified, before hybridization to the ATH1 GeneChip.

(2009) PLoS Genetics 5: e1000621.

G, H mutants



Microgametogenesis (Pollen Development)

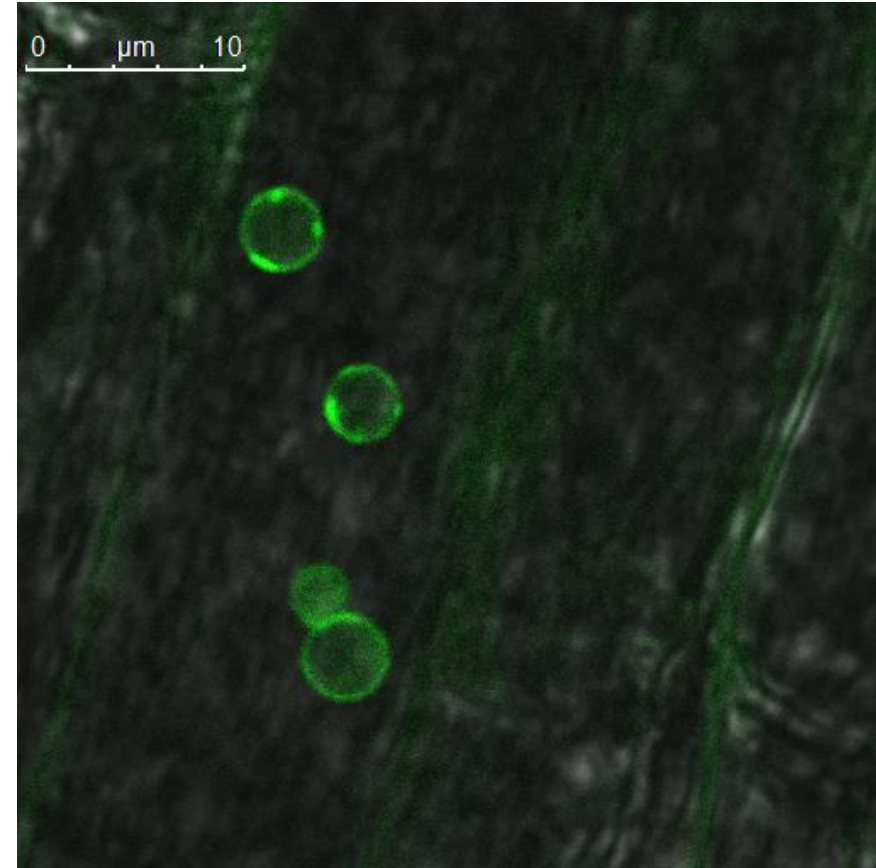
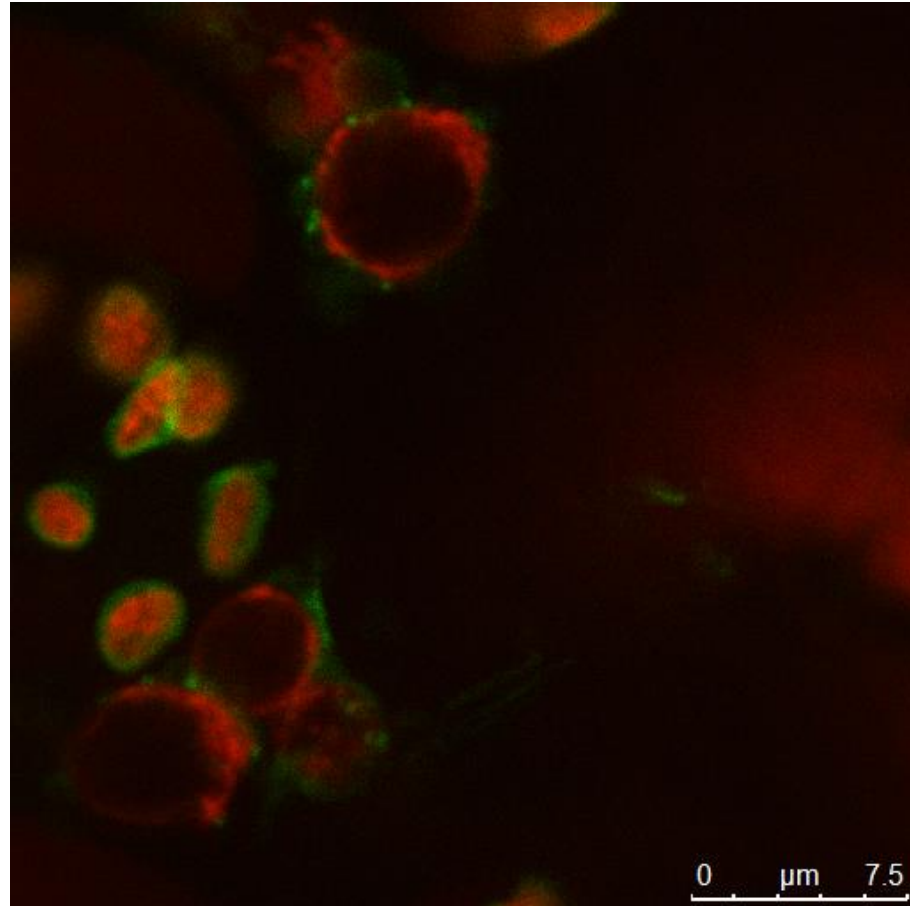


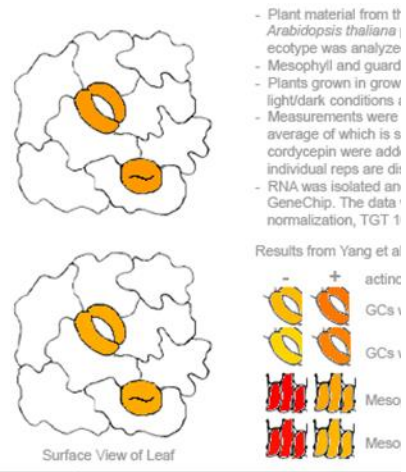
Arabidopsis organellar rhomboid

RBL10-GFP chloroplast
rhomboid → organelle outer
membrane

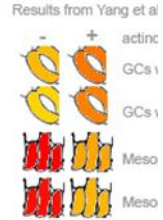
Like other rhomboids,
positioned to activate a protein
within signalling cascade?

Thompson et al. 2012

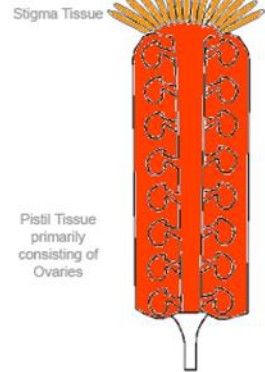




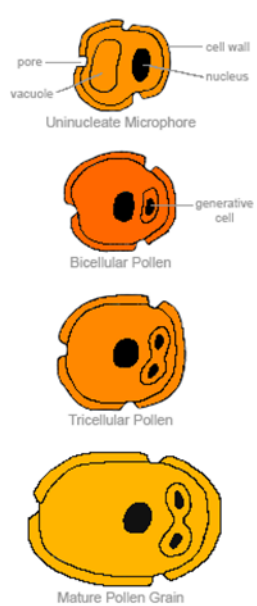
- Plant material from tt *Arabidopsis thaliana* ecotype was analyzed
 - Mesophyll and guard
 - Plants grown in grow light/dark conditions
 - Measurements were average of which is a cordycep were add individual reps are di
 - RNA was isolated an GeneChip. The data normalization, TGT 1



Stigma and Ovaries



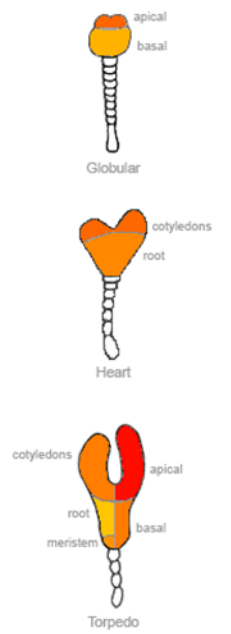
Microgametogenesis (Pollen Development)



- Plant material from the pollen of 5.10 growth stage wild-type *Arabidopsis thaliana* plants of Ler-0 ecotype was analyzed
 - Plants grown under 16/8 hour light/dark conditions at 21°C
 - All measurements were taken in duplicates - the average of which is shown
 - RNA was isolated and hybridized to the ATH1 GeneChip
 - The data were normalized by GCOS normalization, TGT 100

Hony & Twell (2004) *Geno. Biol.* 5:R85

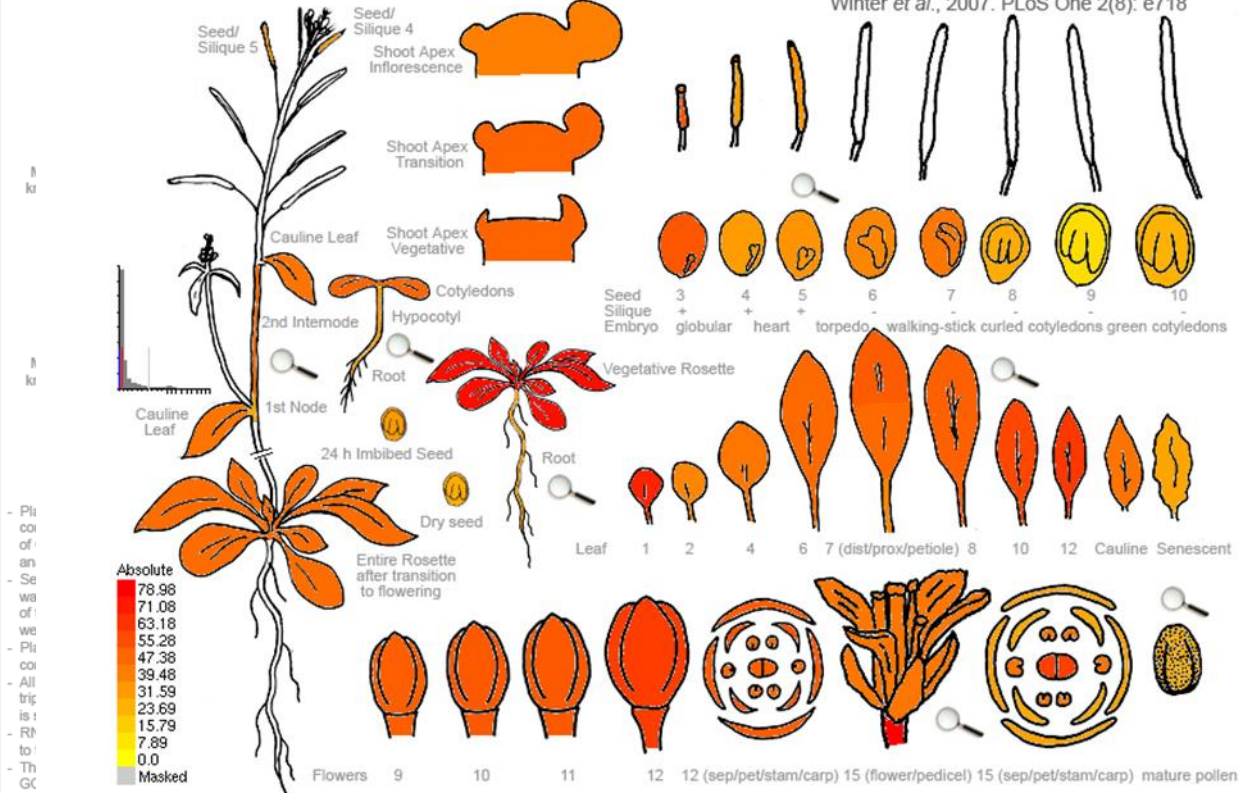
Embryo Development



- Plant material from embryos of wild-type Col-0 *Arabidopsis thaliana* plants of was isolated by laser capture microdissection
 - Plants grown under 16/8 hour light/dark conditions
 - RNA was amplified and hybridized to the ATH1 GeneChip. Note: 3' bias!
 - All measurements were taken in triplicates - the average is shown.
 - Results can be highly variable - standard deviation filtering advisable!
 - The data were normalized by GCOS normalization, TGT 100

Casson et al. (2005) *Plant J.* 42:111

At1g25290 245632_at *ATRBL10*



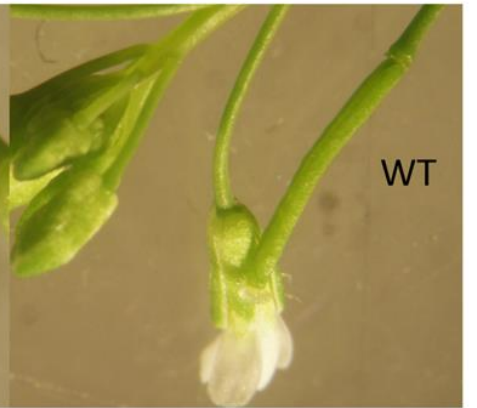
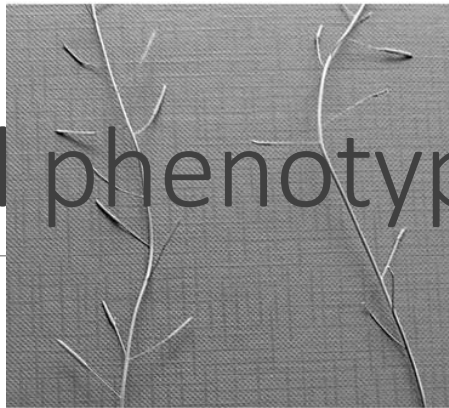
eFP Browser by B. Vinegar, drawn by J. Aills and N. Provart. Data from Gene Expression Map of Arabidopsis Development: Schmid et al., 2005, *Nat. Gen.* 37:501, and the Nambara lab for the imbibed and dry seed stages. Data are normalized by the GCOS method, TGT value of 100. Most tissues were sampled in triplicate.

Chloroplast RBL10 transcription in vegetative and floral tissues

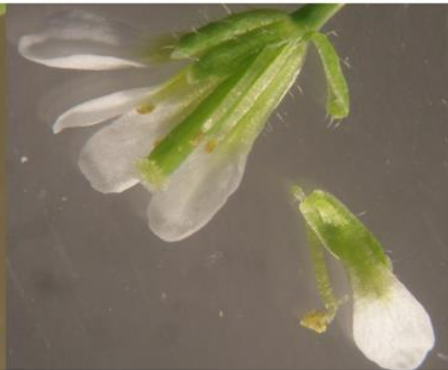
RBL10 floral phenotype

Col0

Mutant



WT



Mut



Comp.
mut.

Do rhomboids often act in concert/related pathways?

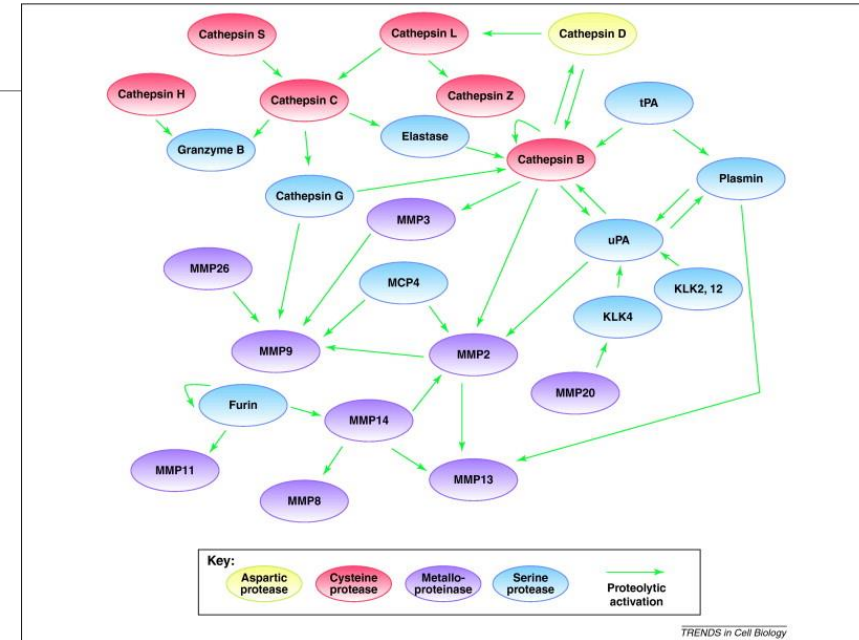
RhmA, B and D (of four 'active' RBDs in Dicty) are in/affect mitochondria

RTPCR suggests overlap

Double A/B mutant is very sick

Literature relates pathways utilising multiple proteolytic events

- Bacterial stress signalling --DegS and YaeL cleaving RseA
- Mitochondrial apoptotic pathways and stress /unfolded protein response
- Higher euk RBDL4 clipping in ER -> proteasome
- Photosystem II repair: DegP/FtsH sequential cleave events
- “members of the small subfamily of type II transmembrane serine proteases ...of particular interest ... compartmentalized expression patterns localizing activity to a limited number of cell types... demonstrated roles as direct contributors to cancer progression”
- “tumor-promoting proteases function as part of an extensive multidirectional network of proteolytic interactions”



Acknowledgements

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Dr John Nichols

Sheffield

Dr Jason King

RHUL

Prof Robin Williams

Dr Grant Otto

Greenwich

Dr Mehak Rafiq

Dr Iskander Ibrahim

Rebecca Vestal

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